

Targeted radioligand therapy: physics and biology, internal dosimetry and other practical aspects during $^{177}\text{Lu}/^{225}\text{Ac}$ treatment in neuroendocrine tumors and metastatic prostate cancer

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Abstract

Radioligand therapy (RLT) has garnered significant attention due to the recent emergence of innovative and effective theranostic agents, which showed promising therapeutic and prognostic results in various cancers. Moreover, understanding the interaction between different types of radiation and biological tissues is essential for optimizing therapeutic interventions. These concepts directly apply to clinical RLTs and play a crucial role in determining the efficacy and toxicity profile of different radiopharmaceuticals.

agents. Personalized dosimetry is a powerful tool that aids in estimating patient-specific absorbed doses in both tumors and normal organs. Dosimetry in RLT is an area of active investigation, as our current understanding of the relationship between absorbed dose and tissue damage is primarily derived from external-beam radiation therapy. Further research is necessary to comprehensively comprehend this relationship in the context of RLTs. In the present review, we present a thorough examination of the involvement of ¹⁷⁷Lu/²²⁵Ac radioisotopes in the induction of direct and indirect DNA damage, as well as their influence on the initiation of DNA repair mechanisms in cancer cells of neuroendocrine tumors and metastatic prostate cancer. Current data indicate that high-energy α -emitter radioisotopes can directly impact DNA structure by causing ionization, leading to the formation of ionized atoms or molecules. This ionization process predominantly leads to the formation of irreparable and intricate double-strand breaks (DSBs). On the other hand, the majority of DNA damage caused by β -emitter radioisotopes is indirect, as it involves the production of free radicals and subsequent chemical reactions. Beta particles themselves can also physically interact with the DNA molecule, resulting in single-strand breaks (SSBs) and potentially reversible DSBs.

Keywords: Radiopharmaceutical therapies, theranostics, DNA damage, dosimetry, ¹⁷⁷Lu, ²²⁵Ac.

Introduction

Nuclear medicine has experienced significant progress in recent years regarding the identification and application of various radiopharmaceuticals for diagnostic and therapeutic purposes. These advancements have been primarily driven by the development of hybrid imaging modalities like SPECT/CT, PET/CT, and PET/MR, which have significantly enhanced the field's capabilities. The evolution of nuclear medicine towards personalized approaches from traditional treatments has been a notable outcome of these technological advancements. The term "Theranostics" derived from the combination of "therapeutic" and "diagnostic," has emerged to describe this integrated approach in current medical practice. Theranostics involves the imaging of tumor cells through the use of a gamma-emitting radiopharmaceutical that targets specific receptors, combined with a therapeutic radionuclide, such as lutetium-177 (¹⁷⁷Lu), yttrium-90 (⁹⁰Y), or Actinium-225 (²²⁵Ac), to effectively eliminate the tumor cells while saving healthy tissues and organs.

This innovative approach represents a significant shift in the treatment paradigm towards more precise and targeted therapies in nuclear medicine [1, 2]. Currently, nuclear theranostics is used in the treatment of neuroendocrine tumors (NETs) with the use of ⁶⁸Ga/¹⁷⁷Lu-DOTA-peptides as well as in prostate carcinoma with ⁶⁸Ga/¹⁸F-PSMA and ¹⁷⁷Lu/²²⁵Ac-PSMA ligands. Individual dose calculation, referred to as dosimetry, is crucial for ensuring the effectiveness and safety of RLT. It is crucial to strike a balance to prevent both under- and over-treatment, while also minimizing toxicity [3-6]. The field of theranostics in the modern era has successfully merged various specialized disciplines such as nuclear medicine, molecular biology, immunology, genomics, radiomics, artificial intelligence (AI), and more [7-9]. This integration has played a crucial role in the advancement of personalized and precision medicine, allowing for a

more cohesive and comprehensive approach to healthcare [10].

This article presents an overview of the core principles of theranostics within the realm of Radioligand therapy, highlighting the interconnection between precision medicine, AI, biology and internal dosimetry in theranostics. A solid grasp of fundamental physics and radiation biology is imperative as a knowledge base to comprehend the variables that impact therapeutic outcomes and the potential toxicities associated with therapy, emphasizing the importance of personalized dosimetry in customizing RLTs for individual patients. Furthermore, a thorough exploration of the utilization of AI in imaging and internal dosimetry within both tumor sites and organs at risk (OARs) is essential, given its pivotal role in forecasting tumor response and overall prognosis in the context of RLT.

This review is designed as a comprehensive, foundational resource in radio-theranostics. Recognizing the multidisciplinary nature of the field, our aim is to provide an integrated overview that is accessible to nuclear medicine technologists, medical physicists, and physicians. For readers seeking a more detailed exploration of specific subtopics, we have included targeted references and subheadings throughout the manuscript that direct to more specialized literature.

Personalized nuclear medicine

Personalized medicine, driven by molecular imaging and theranostics, has emerged as a fundamental approach in tailoring treatment plans to suit the unique needs of each patient. The integration of these advanced technologies has revolutionized the field of medicine, thereby enabling highly precise and effective therapeutic interventions based on individual characteristics and responses [11]. Over the past few decades, there has been a notable transition towards personalized medicine, with the goal of

reducing unnecessary and expensive treatments, while also improving patient care by focusing on better target localization and treatment strategies [11-13]. Diagnostic imaging plays a crucial role in visualizing and localization therapeutic targets, allowing for the identification of specific treatment areas. By utilizing diagnostic scans, healthcare professionals can predict and monitor treatment outcomes, ensuring that the intended sites receive proper treatment during therapy. Additionally, diagnostic imaging aids in determining the most effective treatment strategy.

Tumors are known to display inherent diversity within patients and across different individuals, posing considerable obstacles for targeted cancer treatment [14]. Using a comprehensive whole-body imaging technique is advantageous in this situation, as relying only on a single tumor biopsy may not capture the full diversity of the tumor. This can underestimate the genetic mutational burden, leading to treatment inefficacy or drug resistance [11, 14, 15]. The field of theranostics, which has been in existence for more than seventy-five years, focuses on personalized therapy using molecular imaging.

Within the domain of nuclear medicine physics (NMP), theranostics demands special attention. Moreover, the utilization of SPECT/CT greatly enhances the accuracy and sensitivity of whole-body scintigraphy by offering standardized uptake value (SUV) through meticulous sensitivity calibration. Consequently, this contributes to the progress of personalized medicine [14]. Advancements in theranostics have recently been directed towards neuroendocrine tumors (NETs) by employing ^{177}Lu -DOTA-peptides for therapeutic purposes, alongside ^{68}Ga -DOTA-peptides as PET tracers for diagnostic imaging [15-17] (Figures 1-4).

In addition, the utilization of ^{177}Lu -based SPECT/CT imaging offers several advantages including radiation dosimetry assessment, evaluation of treatment efficacy, and minimizing inadvertent radiation exposure to critical organs such as the kidney, bladder, and red marrow. Specifically for metastatic prostate cancer (mPC), there have been advancements in the development of diagnostic ligands like the ^{68}Ga -PSMA ligands for PET imaging and therapeutic ligands such as ^{177}Lu -PSMA-617 [15, 18] (Figure 5).

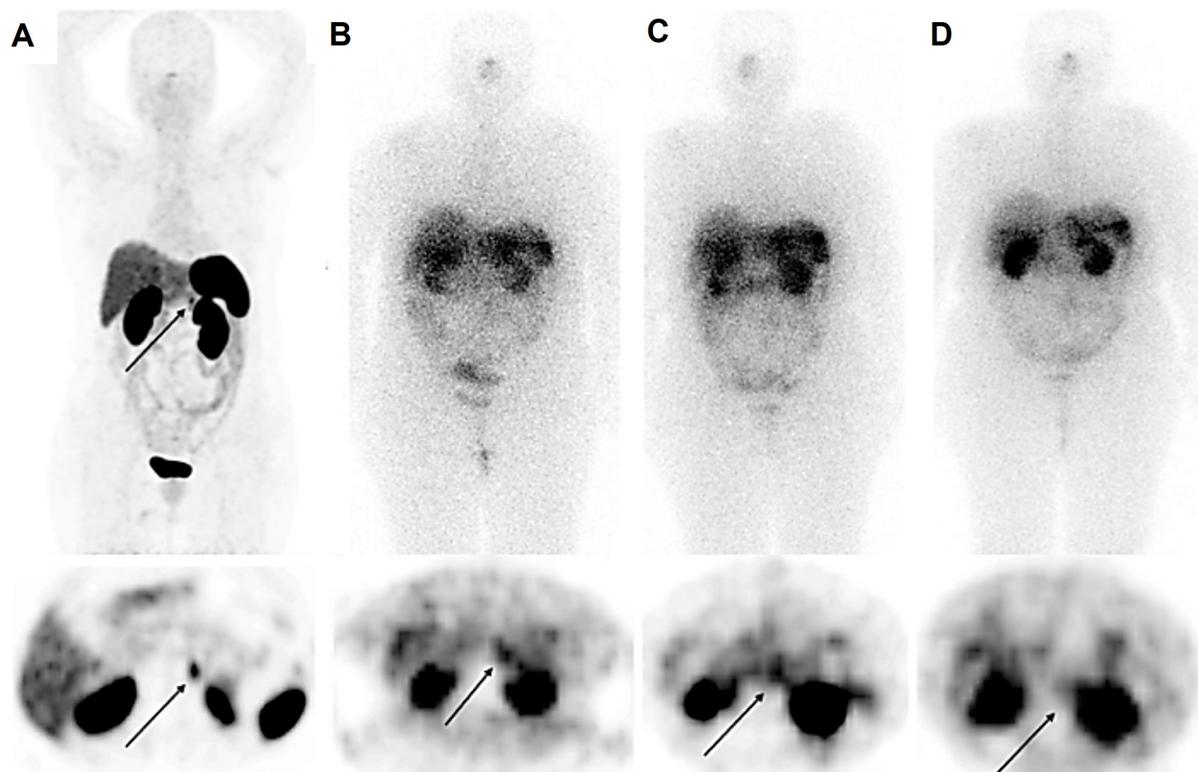


Figure 1. A 56-year-old female with raised calcitonin levels following thyroidectomy, and therefore suspected of recurrent medullary thyroid carcinoma (MTC), underwent ^{68}Ga -DOTATATE PET/CT, which showed radiotracer uptake in the pancreatic tail ($\text{SUV}_{\text{max}} = 10.5$) (A). The patient underwent 3 cycles of PRRT with ^{177}Lu -DOTATATE (10.5 GBq). Post-treatment scintigraphy after the first (B) and second (C) cycles showed an accumulation of radiotracer in the lesions, which was significantly reduced in the third cycle (D), revealing partial response.

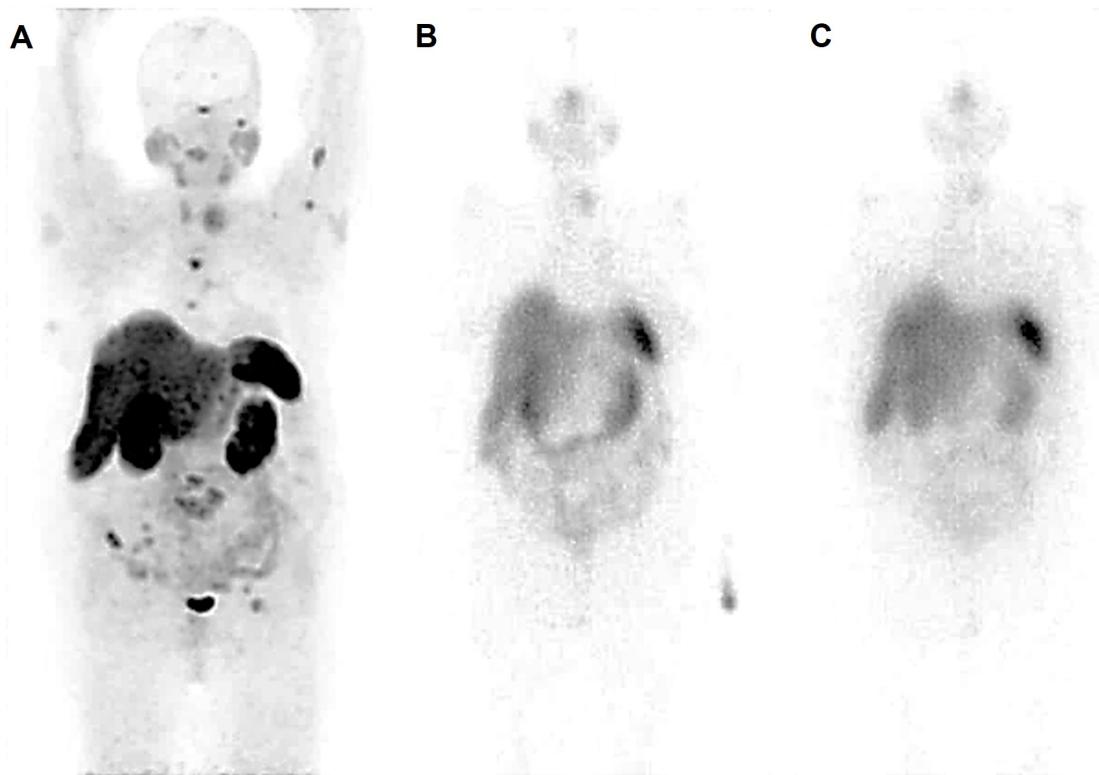


Figure 2. A 70-year-old female presenting with a history of medullary thyroid carcinoma (MTC), raised calcitonin and carcinoembryonic antigen (CEA) levels following thyroidectomy and documented metastatic lesions observed on anatomical imaging, underwent ^{68}Ga -DOTATATE PET-CT to assess the feasibility of RLT, revealing tumoral lesions in the left thyroid lobe as well as pulmonary and multiple bone metastases (A). Hepatic lesions seemed to be non-functional, which could be the result of previous therapies. The patient underwent 2 cycles of PRRT with ^{177}Lu -DOTATATE (cumulative activity, 14.8 GBq). Post-treatment scintigraphy after the first cycle (B) showed radiotracer uptake in the left thyroid lobe, right iliac wing and right sacrum. The second cycle (C) showed an accumulation of radiotracer in the left thyroid region. In follow-up, the levels of calcitonin and CEA increased, indicating progressive disease. Although the hepatic lesions appeared non-functional on the ^{68}Ga -DOTATATE PET/CT scan, potentially due to prior treatments, the presence of multiple metastases throughout the body has been a significant factor in initiating ^{177}Lu -DOTATATE therapy for this patient. Unfortunately, the patient died.

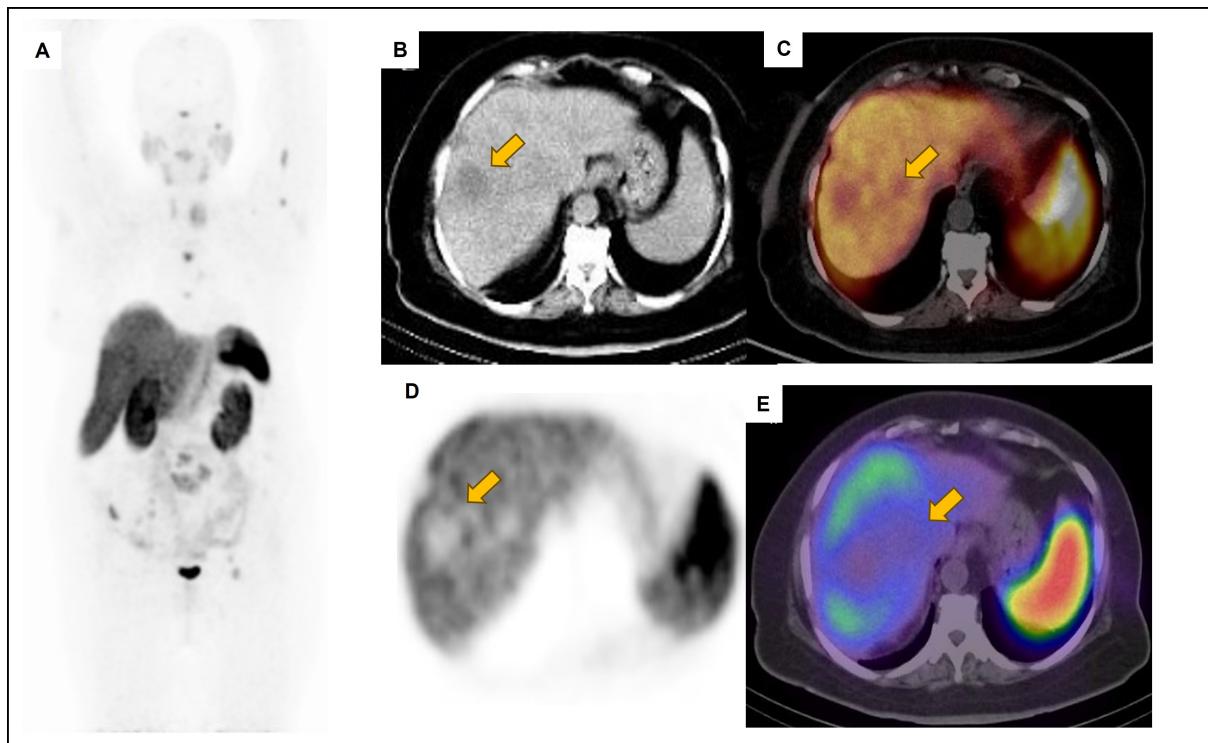


Figure 3. The maximum intensity projection was adjusted to a lower intensity to better highlight the hepatic lesions, which appeared non-functional (potentially due to prior therapies). The patient (same patient shown in Figure 2) subsequently underwent ^{68}Ga -DOTATATE PET/CT (A). Transverse images from CT, PET, and PET/CT fusion are shown in panels B, C, and D, respectively. Non-functional hepatic lesions are further illustrated on the SPECT/CT image (E).

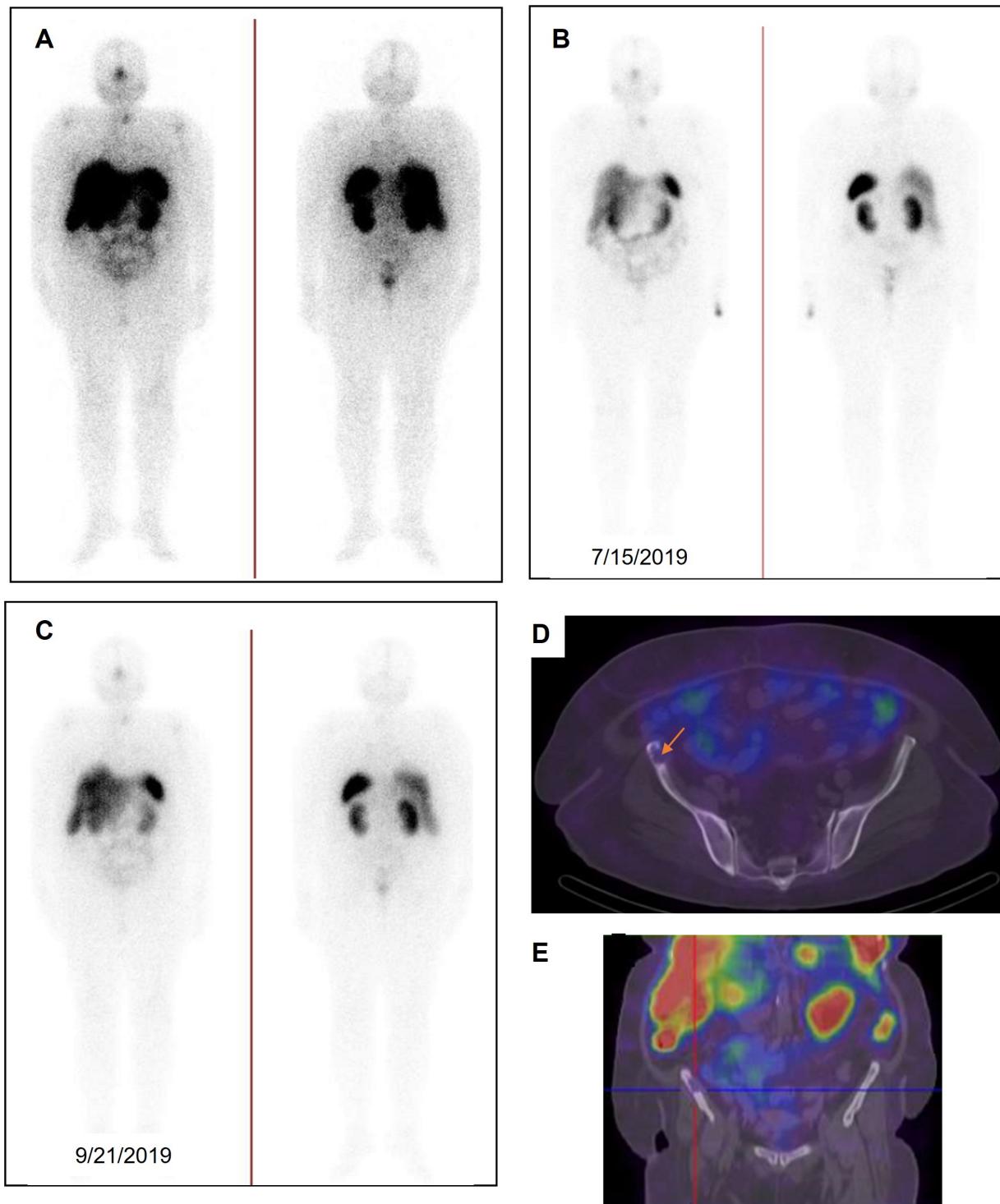


Figure 4. A) Displays higher intensity coloring, indicating a complete response to treatment in the right iliac wing and right sacrum after two cycles of PRRT with ^{177}Lu -DOTATATE, as observed in the baseline PET image (same patient shown in Figures 2 and 3). B) Depicts planar images following the first treatment cycle, while C) represents those after the second cycle. D and E) Present SPECT/CT images to clarify the absence of uptake in the right iliac wing post-treatment, confirming the lack of residual disease activity. The lack of detectable bone lesions in post-treatment images may be due to their small size and the low spatial resolution of ^{177}Lu SPECT scans. We believe that a single therapy cycle usually doesn't yield a therapeutic response, as there hasn't been enough time for the treatment to take effect; thus, these findings represent a baseline assessment at the start of the first cycle.

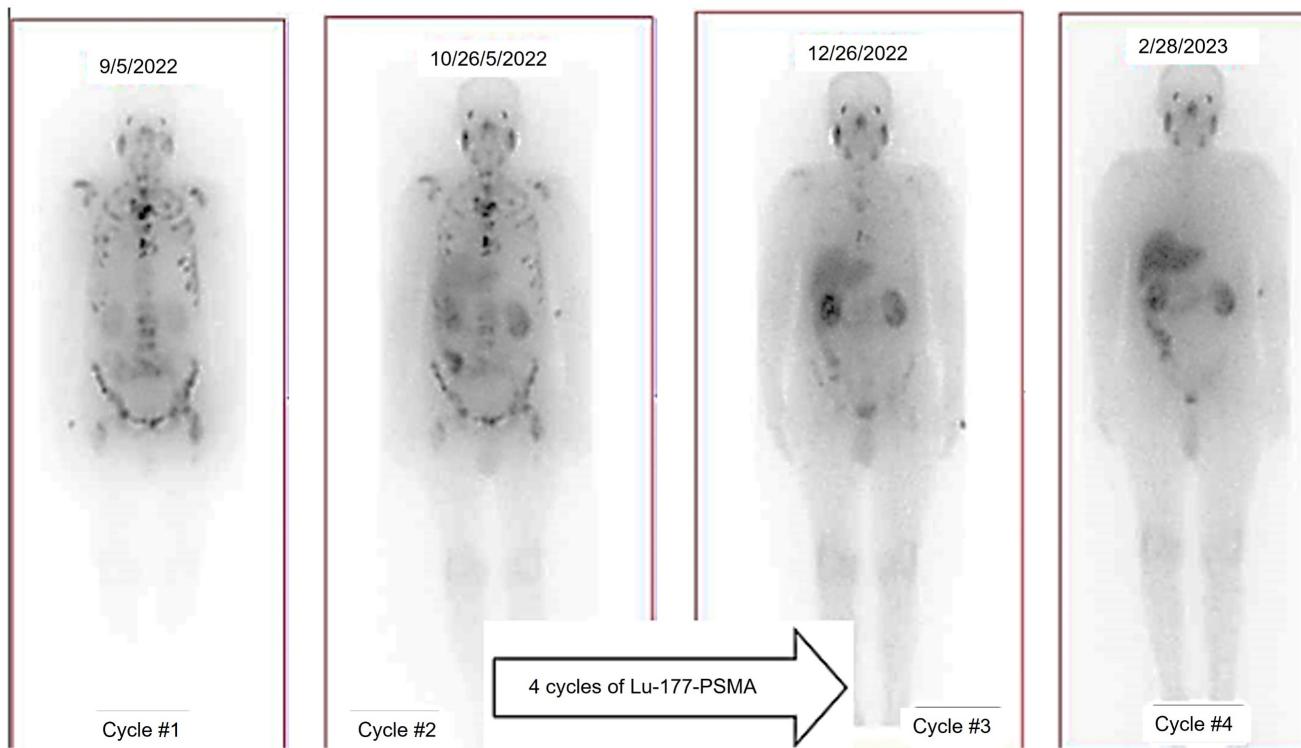


Figure 5. A 69-year-old man with history of radical prostatectomy + pelvic LND (initial PSA=32.0 ng/ml-Gleason score=4+4=8/10 with perineural and lymphovascular invasion). Owing to rising PSA level and widespread bone Mets in previous bone scan, the patient was referred to an oncologist for treatment planning. PSMA PET showed numerous PSMA-avid skeletal lesions throughout the skeleton involving the skull, skull base, sternum, ribs, spine, right scapula, both humeri, pelvic bones, proximal femora with SUV_{max} up to 43.7. Moreover, there are multiple PSMA-avid lymph nodes in the retroperitoneal and pelvic chains (para-aortic, aortocaval, left common iliac, left external iliac, and meso-sigmoid on the left side) with SUV_{max} up to 29.2. After three cycles, in comparison with the previous scans, excellent therapeutic response was noted and uptake in bony structures was reduced. Finally, after the 4th cycle, there is radiotracer uptake in the salivary glands, lacrimal glands, kidneys, bladder and to some extent intestinal lumen as well as liver and spleen (no residual disease is noted; PSA=0.5).

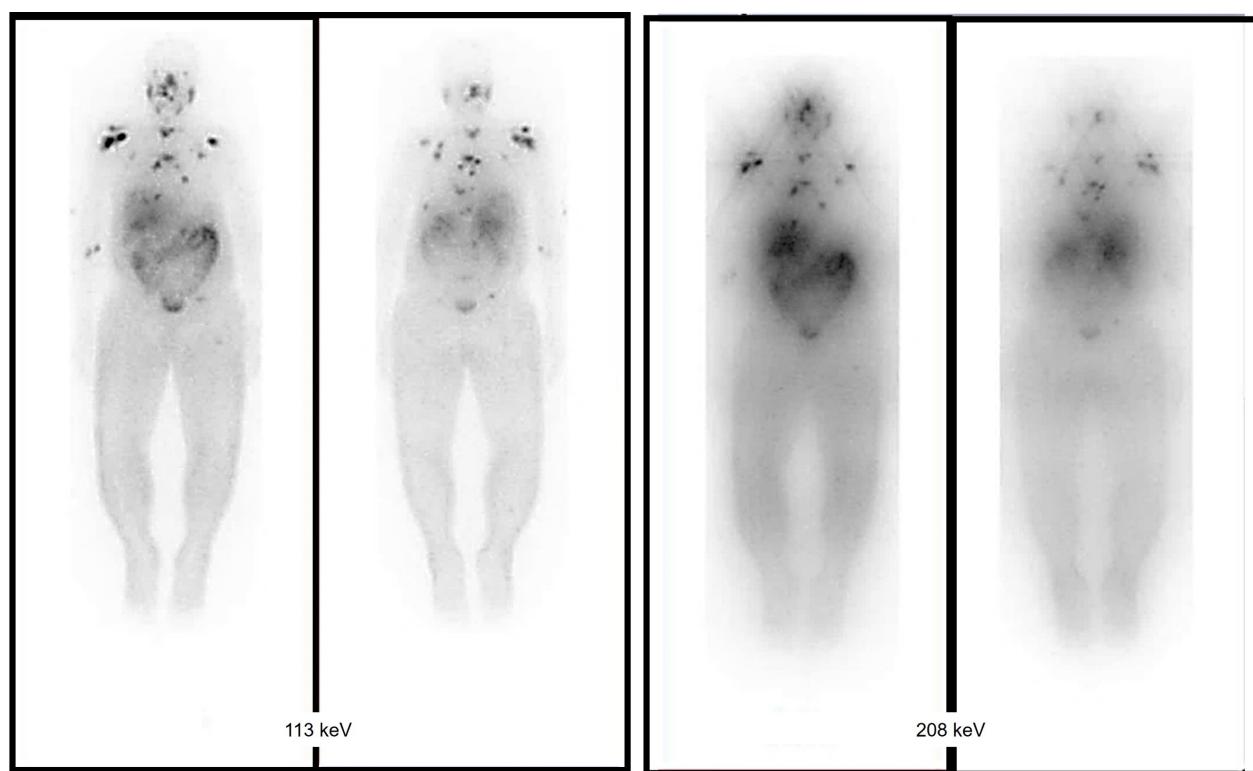


Figure 6. Comparing the effect of choosing the best energy window with using low-energy high resolution (LEHR). A 60-year-old male with known case of prostate adenocarcinoma. Widespread bone metastasis throughout the spine (C6, T1, T8, T9, T10, T11, sacrum), sternum, right femoral head, right clavicle, both scapulae and lateral aspect ribs bilaterally. Acquisitions of these two peaks (113 and 208 keV) using a low-energy high resolution (LEHR) collimator, with the 113 keV peak being regarded as the primary peak, and employing a 20% energy window.

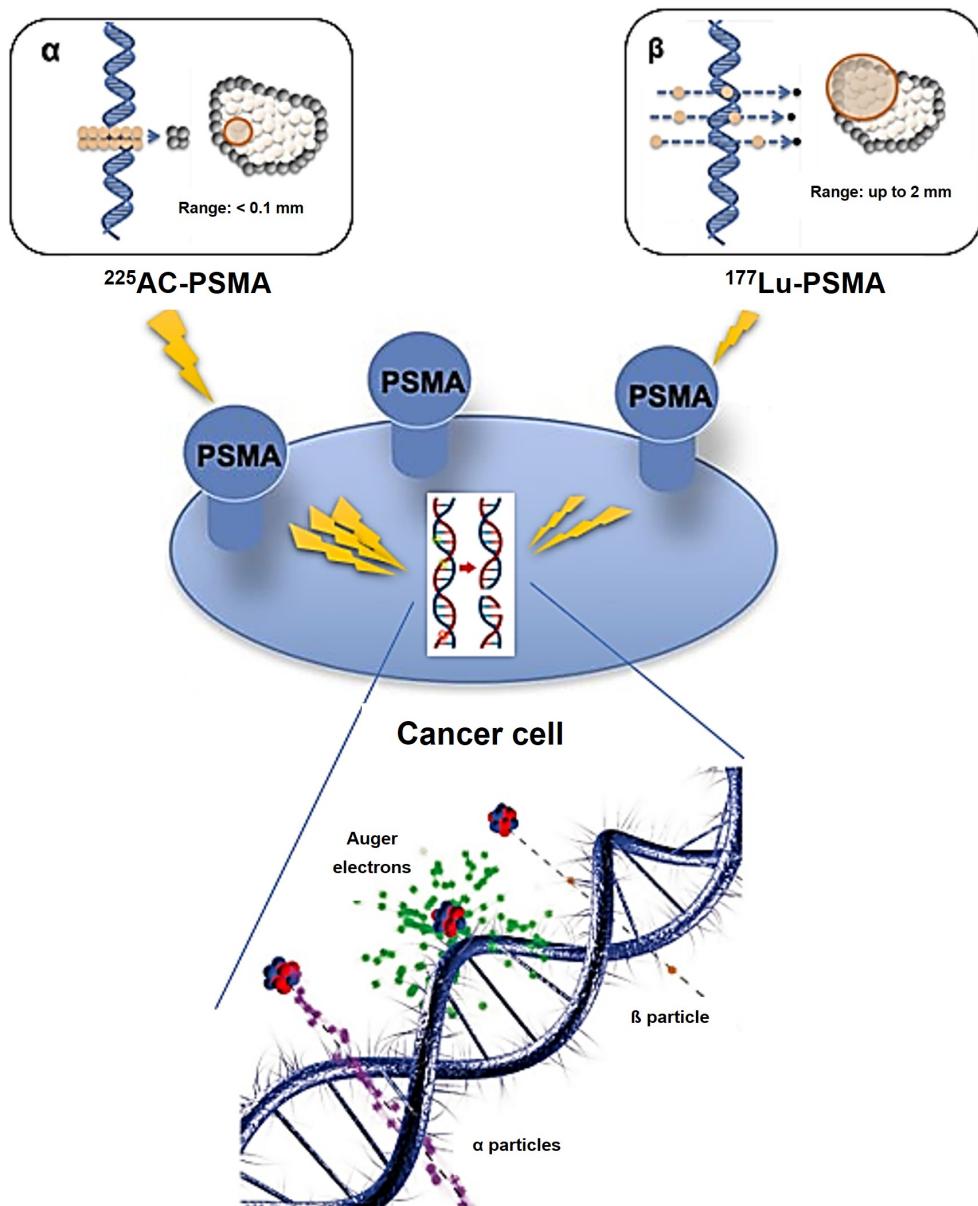


Figure 7. Schematic illustration of the effects of Alpha/Beta particles and their linear energy transfer (LET).

Radionuclides

Alpha vs. Beta particles

Variations in Linear Energy Transfer (LET) and Relative Biological Effectiveness (RBE)

The biological effects of radiation are largely mediated through its interaction with DNA. In particular, alpha particles, due to their high LET, induce dense ionization tracks leading to complex DNA damage, while beta particles, with lower LET, tend to produce more sparse ionization patterns. This difference in ionization density results in distinct patterns of DNA damage and repair mechanisms. β -particles have a low LET of around 0.2 keV/ μ m and

can travel a significant distance of approximately 2-12 mm (equivalent to 20 to 120 cell lengths) [19-21]. The clinically used β -emitting radionuclides, ¹⁷⁷Lu and ⁹⁰Y, are ordered in increasing magnitude according to their maximum energy emission and path length [22]. β -energy of ¹⁷⁷Lu is 0.5 MeV along with two primary gamma-energies of 113 and 208 keV, with respective yields of 6.1% and 10.3%. These gamma emissions facilitate imaging and assessment of radiotracer distribution within the body. Due to scatter (with lower energy) from the 208 keV peak into the 113 keV window, the more intense energy peak was exclusively selected. At the first author's facility, the 113 keV and 208 keV peaks were acquired using a low-energy high-resolution (LEHR) collimator. The

113 keV peak was selected as the primary peak, with a 20% energy window applied, as shown in Figure 6. However, imaging protocols can vary across facilities, depending on equipment, collimators, and energy window settings. For instance, one facility may use a LEHR collimator with a 20% energy window around the 113 keV peak, while others may prioritize the 208 keV peak, employ different collimators, or adjust energy window settings to optimize spectral capture and minimize noise or interference.

Conversely, an α particle bears a positive charge and is approximately three orders of magnitude larger than a β particle. As a result, α particles exhibit a notably higher LET of 80 keV/ μ m in contrast to β particles and cover a considerably shorter distance, usually falling within the span of 50–100 μ m (equivalent to one to three cell lengths) as illustrated in Figure 7 [20].

Theoretically, Auger electron can be used to deliver biological effects to cells. Auger electrons are electrons with low energy that are emitted by radionuclides as a result of decay through electron capture, leading to the ejection of an electron from an electron orbit surrounding the nucleus. These electrons have a restricted travel distance, usually falling within the range of nanometers to micrometers. Similar to α particles, Auger electrons have a high LET and a short range, which enables them to cause localized DNA damage. Despite the limited clinical application of Auger emitters at present, research involving animals and patients treated with Auger emitters has shown promising outcomes, indicating the need for further exploration [23, 24]. In the study conducted by Al-Ibraheem *et al.*, Terbium-161 (^{161}Tb) has emerged as a promising radionuclide for radiotheranostics in different cancer types, such as metastatic castration-resistant prostate cancer (mCRPC). The researchers delve into the potential of ^{161}Tb as an exemplar of auger emission and its ability to rival the effectiveness of ^{177}Lu [25]. Relative biological effectiveness (RBE) distinguishes the biological effects of different radiation types beyond LET. Different types of radiation, like α particles and photons, can result in varying levels of biological damage at the same dose. RBE is calculated as the ratio of the dose required from a standard radiation source to produce an equivalent biological effect to that of test radiation. Factors like dose, dose rate, cell radiosensitivity, repair capabilities, and LET influence RBE. α particles and high LET radiation have a high RBE, indicating they are more effective at causing biological damage compared to β particles or γ emissions (Table 1).

Understanding RBE is crucial for assessing risks and developing radiation protection strategies [26].

The biological outcome of a radiation dose is influenced by the rate at which it is administered. When lower dose rates are used, there is a greater opportunity for repair of sublethal DNA damage compared to high-dose-rate delivery. In contrast to conventional external-beam radiation therapy (EBRT), which delivers a high level of radiation in a short period of time, most RLTs are administered at lower dose rates that decrease exponentially over time. This has important clinical implications for the established toxicity limits for organs at risk (OARs) such as the kidneys, liver, and salivary glands (Figure 8).

Table 1. Characteristics and applications of emitted radiation (α , β -minus and γ radiation).

Type of Emission	LET	Range	Effect	Application
α radiation	+++	+	Dense, highly localized damage	Therapy
β -minus radiation	++	++	Intermediate range and damage	Therapy
γ radiation	+	+++	Sparsely ionizing and long range	Imaging

The current dose limits are primarily based on data from EBRT, and further research is necessary to determine whether higher doses can be tolerated due to the prolonged delivery of radiation in RLTs [3, 27].

Distribution/retention/effective half-Life and administered activity

The duration for a radionuclide to decay to half of its initial activity is known as the physical half-life. On the other hand, the biologic half-life refers to the time it takes to eliminate half of the administered agent through biologic clearance alone. The effective half-life represents the time required for half of the radiotracer activity to clear from the body, accounting for both physical and biological decay. It is important to note that the effective half-life is shorter than both the biologic and physical half-lives due to the combined effects of both decay processes (Figure 9).

Prolonged presence of a substance in a particular tissue is associated with a higher probability of causing harm to that tissue, whether it is a malignant growth or a healthy organ. For example, low extraction fraction of a radiotracer in the bloodstream raises the chances of radiation-induced damage to the bone marrow. A slower elimination rate, leading to an extended biological half-life, increases the risk of bone marrow toxicity from the same dosage administered [28, 29]. Administering a higher activity level can potentially improve the therapeutic impact on specific lesions, but it also increases the potential harm to organs at risk and non-target tissues. In cases where significant toxicity occurs, it is prudent to consider

temporarily or permanently discontinuing the treatment, depending on the severity of the toxicity, to allow for the recovery of organ function. If the patient's organ function improves and they are able to resume treatment, it is generally advised to administer a reduced dose in subsequent cycles to minimize the chances of recurring toxicity. The protocols for managing side effects differ depending

on the specific type of radiotherapy treatment being utilized [30, 31]. The distribution and uptake of the therapeutic agent in tissues contribute to tissue damage, impacting both the antitumor therapeutic effect and organ toxicity. Targeted treatment requires tumor expression of the target (Table 2), and a high level of uptake in tumors is indicative of a more substantial therapeutic response [32-35].

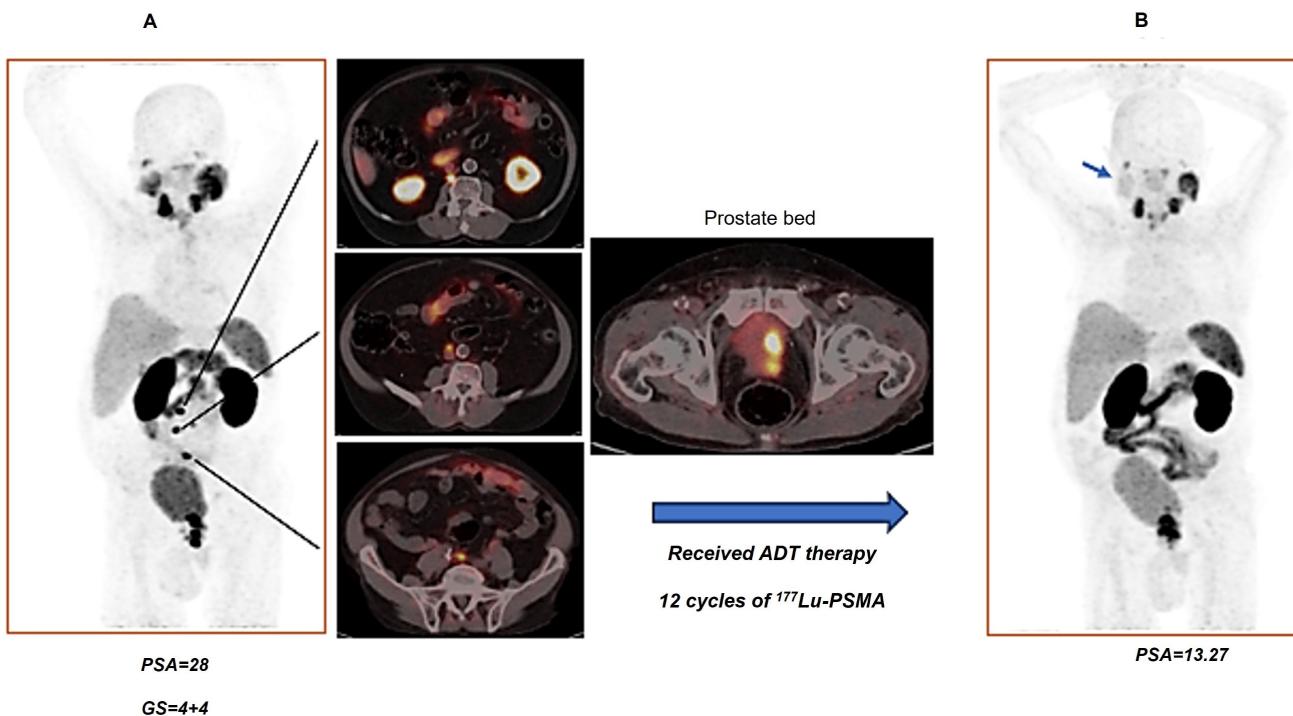


Figure 8. A) 87 years old man with history of prostate adenocarcinoma, underwent hormonal therapy (ADT), referred for recurrence evaluation. A) There are multiple lymph nodes with PSMA uptake in the retrocaval at level L2; ($SUV_{max}=40.87$), precaval ($SUV_{max}=23.59$), paraaortic ($SUV_{max}=9.20$), aortic bifurcation ($SUV_{max}=24.22$) as well as mild PSMA uptake in the left internal iliac lymph node. The prostate gland showed PSMA uptake ($SUV_{max}=42.12$) with invasion to bladder. B) received ADT therapy and 12 cycles of ¹⁷⁷Lu-PSMA up to 2 months before last PSMA PET imaging. Right parotid gland showed decreased physiologic tracer uptake comparing with left parotid gland (because of ¹⁷⁷Lu-PSMA therapy- blue arrow in the right image). All retroperitoneal and paraaortic LNs which is mentioned in previous scan showed no more uptake. Intense increased PSMA uptake in the prostate gland, involving a significant portion of the gland, with extension to the seminal vesicles and invasion of the posterior wall of the urinary bladder. Comparison with the previous scan showed progression. Findings are consistent with partial response to therapy.

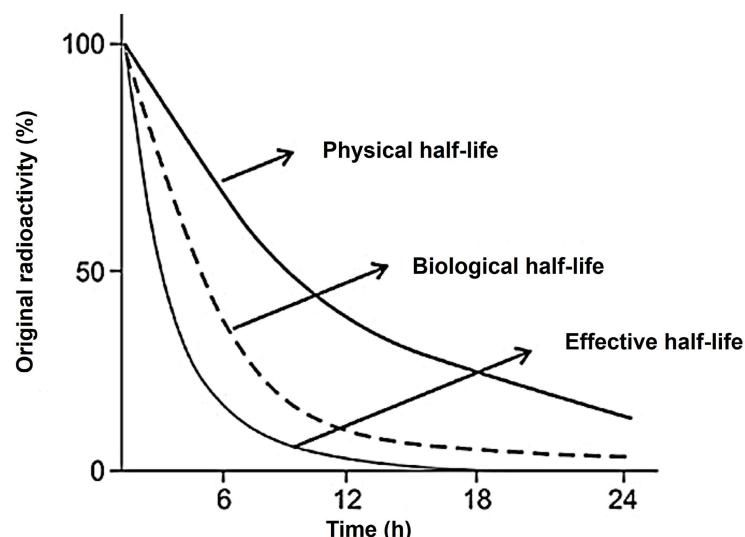


Figure 9. The effective half-life, accounting for both physical and biologic decay, is the time for half of the radiotracer activity to clear from the body and is consequently shorter than both the biologic and physical half-lives.

Table 2. PSMA expression score and eligibility for RPT according to the PROMISE V2 criteria.

Score	Uptake Relative to Internal Reference	Eligibility for RPT
0	≤Blood pool	No
1	≤Liver and > blood pool	No
2	≤Parotid gland and > liver	Yes
3	>Parotid gland	Yes

Organs that exhibit elevated physiologic uptake are more susceptible to the harmful effects of radiation. An example of this is the occurrence of dry mouth, known as xerostomia, which was reported as a prevalent side effect in 38.8% of patients undergoing ¹⁷⁷Lu-PSMA ligand treatments in the VISION trial (Figure 10) [32, 36].

Table 3. Example of emerging molecular targets in clinical trials (NET).

Molecular Targeting Mechanism	Trial Registration No. and Agents	Type of Tumor
	NCT02609737: ⁶⁸ Ga-DOTA-JR11/ ¹⁷⁷ Lu-DOTA-JR11	NET
	NCT02592707: ⁶⁸ Ga-OPS202/ ¹⁷⁷ Lu-OPS201	NET
Somatostatin receptor antagonist	NCT04997317: ¹⁷⁷ Lu-satoreotide	Meningiomas
	NCT05017662: ¹⁷⁷ Lu-IPN01072	NET, long-term surveillance for secondary malignancies
	NCT05359146: ¹⁶¹ Tb-DOTA-LM3	NET

Radionuclide Therapy

Current Clinical Practice and Future Directions

Somatostatin Receptor–targeted Radionuclide Therapy

NETs in the pancreas, lung and midgut that exhibit somatostatin receptors (SSRs) expression, in particular the subtype 2 (SSR2a), can be identified through the utilization of ⁶⁸Ga-DOTA peptides such as DOTATATE, DOTANOC, and DOTATOC. A theranostic strategy involving ¹⁷⁷Lu-DOTA-peptides or ⁹⁰Y-octreotate can be employed for the treatment of these tumors [37]. During the critical third phase of the NETTER-1 trial, individuals diagnosed with advanced midgut NETs who received treatment with ¹⁷⁷Lu-DOTATATE experienced a notable rise in progression-free survival (PFS) after 20 months in contrast to the control cohort (65.2% versus 10.8%, respectively). Furthermore, a significantly increased tumor response rate was noted [38]. The NETTER-1 trial, unfortunately, did not show a statistically significant improvement in overall survival (OS) after 5 years. This lack of significance can be attributed in part to a substantial crossover rate within the control group, with 36% of participants eventually undergoing SSR-coupled radionuclide therapy [39]. Consequently, significant clinical guidelines integrated this treatment into their protocols. Moreover, the NETTER-1 study illustrated that patients treated with ¹⁷⁷Lu-DOTATATE encountered a superior quality of life across various domains such as overall health, body image, functionality (both general and occupational), diarrhea, pain, fatigue, and concern about the illness. Further investigation and advancement in the field of RLT is imperative for

patients diagnosed with advanced midgut NETs. A recent phase 2 clinical trial focused on exploring the efficacy of an SSR-targeted α -particle therapy utilizing ²²⁵Ac-DOTATATE [40]. The results of this trial showcased promising responses and PFS outcomes specifically for gastroenteropancreatic neuroendocrine tumors (GEP-NET). Previous preclinical and initial clinical research has suggested that SSR2 antagonists may exhibit a higher receptor binding density, leading to a more favorable tumor-to-background ratio (TBR) and improved lesion detection when compared to agonists [41, 42]. The heightened receptor binding capacity of antagonists has the potential to increase sensitivity, enabling the treatment of tumors with lower receptor density. Table 3 presents a comprehensive overview of five ongoing trials that employ investigational theranostic pairs based on SSR antagonists for SSR-expressing NETs and meningiomas.

This highlights the active and innovative nature of RLT in this field. Furthermore, there is a growing interest in investigating the use of SSR-coupled RLT at an earlier stage in the treatment process. This is exemplified by the NETTER-2 trial, which aims to evaluate the efficacy of ¹⁷⁷Lu-DOTATATE as a first-line treatment for untreated patients with metastatic grade 2–3 NETs. The trial involves randomization of patients to receive either ¹⁷⁷Lu-DOTATATE in combination with long-acting octreotide or high-dose long-acting octreotide alone.

Prostate-specific Membrane Antigen–targeted Radionuclide Therapy

In the TheraP trial's second phase, the utilization of ¹⁷⁷Lu-PSMA-617 demonstrated a more remarkable decrease in prostate-specific antigen (PSA) levels when compared to cabazitaxel for patients with

metastatic castration-resistant prostate cancer (mCRPC) that was progressing [43]. The phase 3 VISION trial (ClinicalTrials.gov no. NCT03511664) revealed additional results indicating that individuals who underwent treatment with ^{177}Lu -PSMA-617 demonstrated improved PFS as observed through imaging, as well as a greater median OS (15.3 vs. 11.3

months, respectively) in comparison to patients who solely received standard-of-care treatment [36]. The FDA has recently approved ^{177}Lu -PSMA-617 for the treatment of mCRPC, in addition to the utilization of ^{68}Ga -PSMA-11 PET for diagnostic imaging purposes (Figure 11).

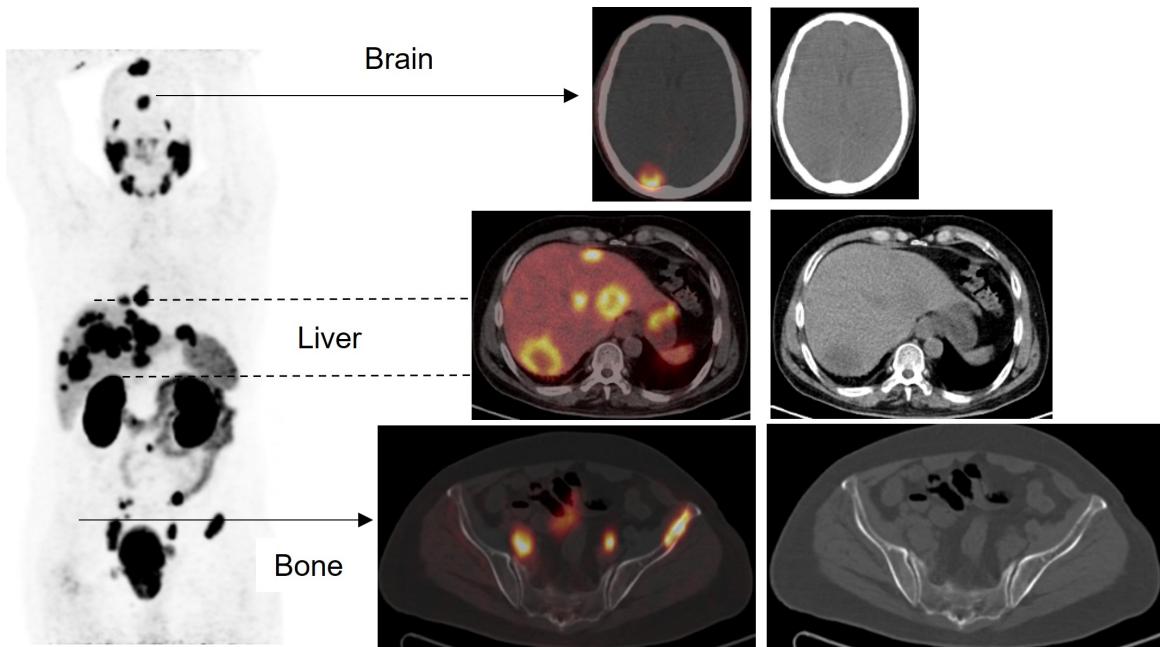


Figure 10. A new case of prostate cancer (poorly differentiated carcinoma) with a PSA level of 21 has been referred for staging. The staging evaluation revealed the presence of multiple lymph nodes with PSMA uptake in the para-aortic, common iliac, external, and internal iliac chains, measuring up to 28mm with an SUV_{max} of 15.10. Additionally, the liver exhibited multiple PSMA-avid masses in both lobes, measuring up to 52mm with an SUV_{max} of 16.16. The prostate gland appeared enlarged and lobulated, displaying significant inhomogeneous PSMA uptake. There is a likelihood of invasion into the infero-posterior wall of the urinary bladder, with an SUV_{max} of 18.07. Metastases were observed in various locations, including the skeletal system, both liver lobes, bilateral lung field, and brain. Notably, there was intense uptake in the salivary glands.

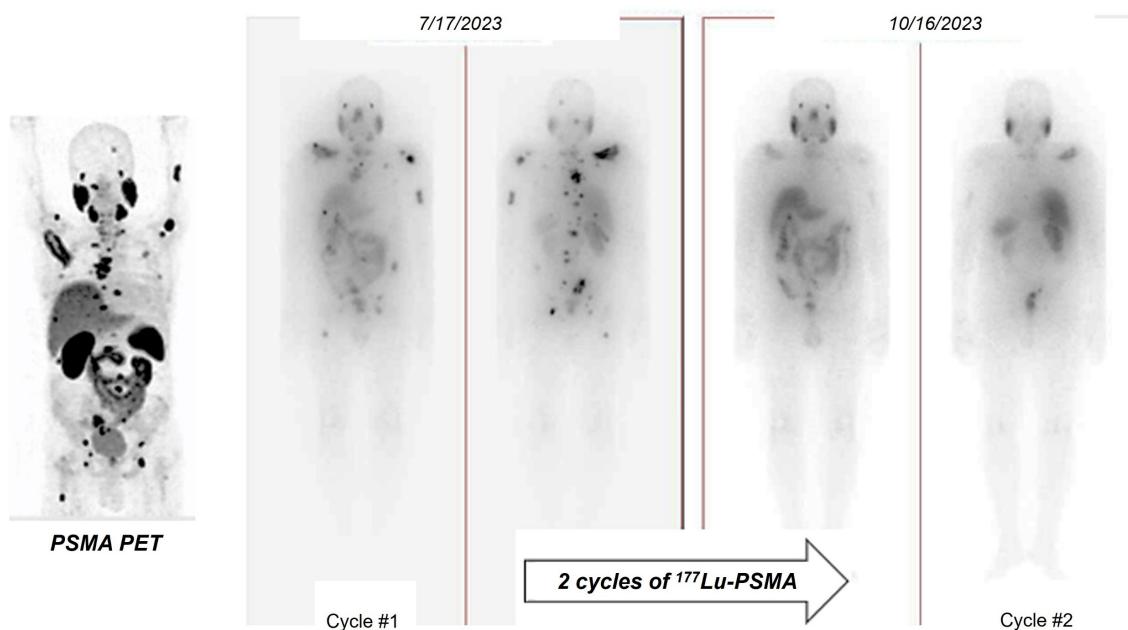


Figure 11. A 63-year-old man with history of prostate adenocarcinoma (GS=4+3=7/10 and bone mets) underwent the chemotherapy and Abiraterone with increasing the PSA level to 135 ng/ml. The scan showed increased tracer uptake in the right scapula, right iliac and sacrum. Physiologic tracer uptake was seen throughout salivary gland, liver, GI tract and urinary bladder. Metastatic involvement in the right scapula, right iliac and sacrum which is comparing with previous scan showed partial response to treatment.

¹⁸F-DCFPyL PET has been utilized in clinical trials as the diagnostic pair targeting PSMA [44, 45]. The realm of PSMA-coupled RLT is currently experiencing significant clinical advancements, concentrating on the advancement of targeted α -particle therapy in conjunction with PSMA ligands. A groundbreaking study in 2016 showcased an impressive treatment outcome through the utilization of ²²⁵Ac-labeled PSMA ligands (²²⁵Ac-PSMA-617) in mCRPC patients [46]. Additional research has provided further evidence of the effectiveness and safety of ²²⁵Ac-PSMA-617 as a viable treatment choice. A comprehensive analysis and evaluation of 256 patients who underwent treatment with ²²⁵Ac-PSMA-617 revealed an impressive overall biochemical response rate of 62.8%. Furthermore, a molecular response rate of 74% was observed through the use of ⁶⁸Ga-PSMA-11 PET/CT scans. The median estimates for PFS and OS were determined to be 9.1 months and 12.8 months, respectively [47]. Additional randomized controlled trials in the future are essential to further establish the effectiveness of therapy and the benefits it provides for survival. Currently, phase 1 and 2 trials, as well as registry data, are actively enrolling patients. Despite the increased risk of xerostomia, ²²⁵Ac is actively being studied in prospective clinical trials. An ongoing phase 2/3 trial (NCT06402331) is underway, with additional trials anticipated to commence soon, potentially paving the way for future FDA approval.

The potential application of this approach for eliminating the primary tumor site is currently being investigated in clinical studies, including the recently completed LuTectomy trial (ClinicalTrials.gov registration no. NCT04430192). These trials will provide valuable insights into the feasibility of utilizing targeted treatment in earlier stages of the disease.

To provide a robust foundation for current clinical practices, this section included an expanded discussion on recent clinical trials. Emphasis has been placed on methodological nuances, such as trial design, patient selection criteria, and outcome evaluation. Detailed case studies and data analyses have been incorporated to illustrate these points, with further reading provided in [48].

Biology and DNA damage

Recent advances in radiobiology have significantly enhanced our understanding of the molecular and cellular responses to radiation. In this section, we elaborate on key mechanisms, such as DNA damage response, cell cycle checkpoints, and repair pathways. For further detailed insights, readers are encouraged to consult the extensive reviews cited

herein [49, 50].

Cellular Repair Mechanisms

DNA is a pivotal molecule that preserves essential genetic information necessary for the growth and survival of organisms. It is renowned for its relatively superior stability when compared to other biological compounds [51, 52]. Although DNA is stable, it can be modified by internal and external factors, leading to harmful mutations in cells [53]. Various factors, like replication errors, mismatched DNA bases, spontaneous deamination, and oxidative damage from reactive oxygen species (ROS), can cause DNA damage [54, 55]. DNA damage can result from external factors like UV radiation, chemicals, and other environmental elements [52, 56] (Figure 12).

External factors can cause DNA single-strand breaks (SSB) or double-strand breaks (DSB). SSB is when one DNA strand breaks, while DSB is when both strands break [57] (Figure 13). In cancer, disruptions in DNA damage response can induce mutations and instability, driving disease progression [53, 58, 59]. Tumors grow quickly due to unstable genes, which can be treated with chemotherapy, radiation therapy, or radiopharmaceuticals [53, 60]. It is crucial to understand the effects of targeted radiation therapy on tumors and minimize unintended harm by studying DNA damage and repair mechanisms triggered by different radioisotopes and molecular targets [56, 59].

High-LET particles and its effects on DNA damage

Alpha-particle emitters. The use of α -particle emitters in targeted therapeutics is gaining attention due to their high energy levels (4-8 MeV) and limited emission range in tissues. Using a targeted radiopharmaceutical labeled with an α -emitter allows for precise delivery of radiation to tumors while minimizing exposure to healthy tissues [61-63]. Research suggests that α -emitting radioisotopes, with their high energy and charge, can directly interact with DNA, leading to the formation of irreversible DSBs within the DNA structure [64]. α -emitters cause direct DNA damage and can also generate free radicals, leading to some indirect DNA damage [65]. The distinction between direct and indirect DNA damage and their long-term clinical effects is unclear and requires further investigation in future research.

Low-LET particles and its effects on DNA damage

Beta-particle emitters. The main connection between β -emitters' toxicity in mammalian cells and their effects is the indirect DNA damage caused by

ROS and oxidative stress [66, 67]. These mechanisms primarily create SSBs, with DSBs less commonly found in affected cells [59, 68]. Research shows that β -emitters, such as ^{177}Lu , primarily work by creating SSBs for therapeutic purposes. While tumor cells can repair these breaks, the accumulation of damage can overpower repair processes, leading to cell death and tumor reduction [69]. However, the use of ^{177}Lu -DOTATATE or ^{177}Lu -PSMA ligands [70, 71] has led to an increase in DSBs with higher doses. Research has been done to study how ^{177}Lu , a common

β -emitter, affects cellular death mechanisms [72-74]. Beta-emitters moving through biological tissue primarily cause DNA damage indirectly by generating free radicals like ROS, leading to chemical reactions. These processes can result in early or late apoptosis, mutations, and genomic instability [75]. β -particles can cause direct DNA damage by interacting with DNA, displacing electrons and causing ionization. This can lead to SSBs and potentially reversible DSBs [76, 77].

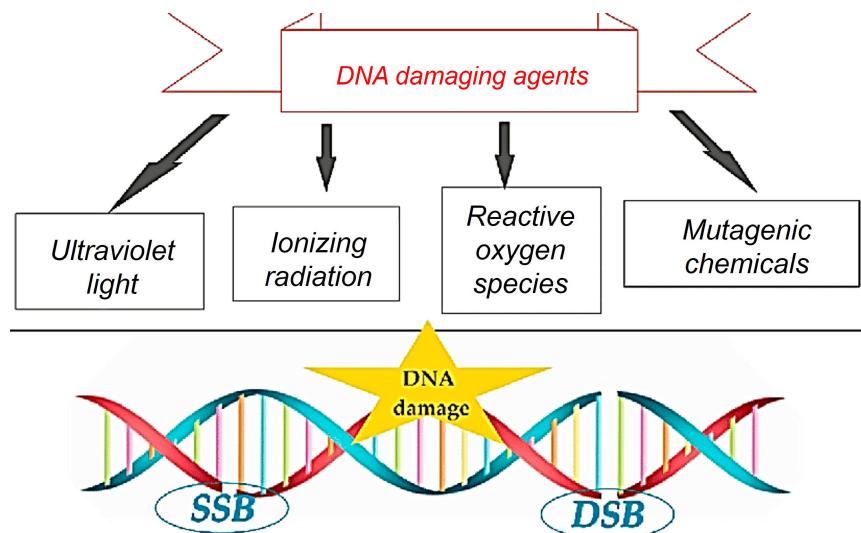


Figure 12. DNA damage is caused by external factors such as environmental, physical, and chemical agents like ultraviolet and ionizing radiation, alkylating agents, and crosslinking agents.

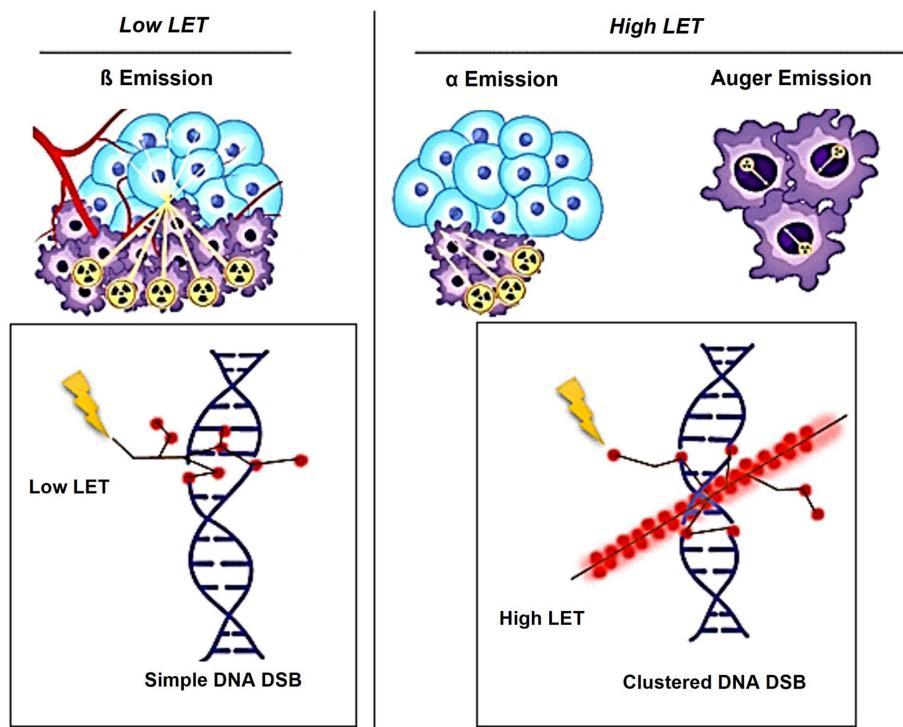


Figure 13. These external factors can lead to DNA single-strand breaks (SSB) or double-strand breaks (DSB), where SSB involves the breakage of one DNA strand while the other remains intact, and DSB involves the breakage of both DNA strands.

DNA damage repair pathways in the cellular response to Beta/Alpha particle emitters

Cancer cells lack DNA repair pathways, making them susceptible to radiotherapy and DNA-damaging substances. EBRT and RLT exploit this vulnerability, causing irreparable DNA damage and leading to the death of cancer cells [56, 59]. An investigation found a link between DNA damage response irregularities and increased PSMA expression in prostate cancer patients, potentially improving response to PSMA-targeted RLT [78]. In the context of ²²⁵Ac-PSMA-617 treatment, two individuals with mCRPC and BRCA1 gene mutations showed longer survival compared to patients without DNA damage response (DDR) mutations [79]. Radioimmunotherapy using α -emitters is less likely to cause resistance in cancer cells compared to beta-emitters, possibly due to the creation of permanent double-strand breaks in DNA [80, 81]. However, resistance to α -emitters can be attributed to various DDR mechanisms and signaling pathways within cancer cells [82-84]. The selection of a repair pathway for DNA DSBs is complex and influenced by factors, such as the number and type of DSBs [81].

Impact of dose rate on DNA damage and repair

Dosimetry in RPT lacks a specific definition found in established EBRT protocols, making it difficult to accurately calculate and understand absorbed doses and their dispersion in specific tissues or organs [85]. Determining the dose rate (DR) is crucial for understanding radiation's impact on cellular structures, gene expression, cellular responses, and cell death mechanisms. Radiopharmaceuticals release radiation gradually, resulting in a fluctuating and declining dose rate. The energy and distribution of the dose depend on the radionuclide used. Factors like physical half-life, specific activity, biological half-life, and cell repair ability influence the DR in radiation therapy. Lower dose rates cause less harm and dispersed radiation, leading to repairable sublethal damage [85, 86]. Researchers have found that repair foci increase non-linearly at low dose rates, suggesting repair mechanisms may be more effective at low doses than high doses [87]. Contradictory findings challenge the idea that low levels of radiation do not activate genes necessary for DNA repair and only result in minimal or no repair of DNA damage [88, 89]. Studies suggest that DNA repair is limited at low dose rates due to insufficient DNA damage initiation, leading to inadequate activation of repair genes. This disparity in cellular response to double-strand breaks is evident

when comparing low and high doses of low LET radiation [88, 90-92]. Lower doses may increase the risk of cancer more than estimated based on higher doses using linear extrapolation due to the absence of repair mechanisms [89, 93]. The bystander effect shows how indirect harm can result from exposure to RLT [94, 95]. The spread of damage from irradiated cells to nearby non-irradiated cells is significant [96, 97]. Therefore, it is reasonable to suggest that bystander signaling may have been less effectively induced at the higher dose rate compared to the lower dose rate. Understanding the effects of different radioisotopes on DNA damage and repair processes, estimating absorbed doses through dosimetry, comparing radioisotopes with varying energy and emission profiles, and evaluating photonics versus electronic emissions are all crucial for advancing RLT. Enhancing preclinical models, studying RLT effects on the tumor microenvironment, exploring combination treatments, and assessing short and long-term toxicities are also essential. Collaboration among oncologists, nuclear medicine specialists, radiation oncologists, physicists, and biologists is necessary to bridge this knowledge gap.

Dosimetry

Investigational uses of dosimetry in radionuclide therapy

In RLT, individual variations in peptide pharmacokinetics among patients necessitate personalized treatment strategies, such as adjusting the number of treatment cycles or the amount of administered activity [98]. One approach to therapy planning involves determining the maximum tolerable absorbed dose to non-target organs, known as the "as high as safely attainable" (AHASA) approach [99], as opposed to the traditional "as low as reasonably achievable" (ALARA) approach, which aims to minimize radiation exposure to non-target tissues. Despite this, the current practice typically involves administering a fixed activity of 7.4 GBq per cycle, as seen in the NETTER-1 trial. As a result, personalized dosimetry is often conducted primarily to ensure safety, and assess the absorbed dose to the tumor, rather than to optimize the administered activity, and evaluate the dose-response relationship. Accurate estimation of the activity in the targeted organs at multiple time points is essential for patient dosimetry [100, 101]. Hence, the preliminary calibration and quantification steps play a crucial role [102, 103]. Despite the existence of Medical Internal Radiation Dose (MIRD) guidelines [104, 105], there is a pressing need for a standardized dosimetry protocol to assess safety, and toxicity, and conduct dosimetric

evaluations. According to a recent review by Huizing *et al.* [106], dosimetry in RLT is not commonly practiced due to various challenges in

implementation, the time-consuming nature of non-standard dosimetry methods, and the lack of supporting evidence in the literature.

Table 4. Software packages for phantom-based dosimetry.

Name	Availability	Decay Data	Number of Radionuclides	Phantoms	Specific Organ Models
OLINDA/EXM 1 (2004) Olinda/EXM® 2.0	Distributed by Vanderbilt University, presently Withdrawn from the market	RADAR website	Over 800	Cristy and Eckerman + pregnant female series	Peritoneal cavity, prostate gland, head and brain, kidney and spheres
Organ DosimetryTM with	Distributed by Hermes Medical	RADAR website	Over 1000	RADAR phantoms + animal phantoms	Peritoneal cavity, prostate gland, head and brain, kidney and spheres
IDAC 2.1 (2017)	Free	ICRP 107	1252	ICRP 110	Spheres
3D-RD-S (2020)	Distributed by Rapid, LLC	ICRP 107	1252	ICRP 110 and ICRP 143	Spheres
MIRDcalc (2021)	Free	ICRP 107	333	ICRP 110, ICRP 143 and weight-based phantoms	Spheres

Table 5. Main commercial software packages for patient-specific dosimetry.

Name	Manufacturer	Dose Conversion Method	Supported Therapy Radionuclides	CE/FDA Approval
SurePlan™ MRT	MIM Software Inc.	VSV	¹⁷⁷ Lu, ¹³¹ I	CE/FDA
Planet® Dose	DOSIsoft	VSV/LD M	¹⁷⁷ Lu, ¹³¹ I	CE/FDA
Voxel DosimetryTM	Hermes Medical solutions	Semi-MC	⁶⁸ Ga, ¹²³ I, ¹³¹ I, ¹¹¹ In, ¹⁷⁷ Lu, ^{99m} Tc, ⁹⁰ Y, ⁸⁹ Zr, ²²³ Ra, ¹⁶⁶ Ho	CE/FDA
QDOSE®	ABX-CRO	VSV	¹¹³ C, ¹⁵ O, ¹⁸ F, ⁴⁴ Sc, ⁶⁴ Cu, ⁶⁸ Ga, ⁸⁶ Y, ⁸⁹ Zr, ⁹⁰ Y, ¹²⁴ I, ⁸⁹ Sr, ^{99m} Tc, ¹¹¹ In, ¹³¹ I, ¹⁵³ Sm, ¹⁶⁶ Ho, ¹⁷⁷ Lu, ¹⁸⁶ Re, ¹⁸⁸ Re	CE
SurePlan™ LiverY90	MIM Software Inc.	VSV/LD M	⁹⁰ Y microspheres	CE/FDA
Planet®	DOSIsoft	Dose VSV/LD M	⁹⁰ Y microspheres	CE/FDA
Hybrid3DTM SIRT	Hermes Medical solutions	LDM	⁹⁰ Y microspheres	CE/FDA
Simplicit90YT M v2.4	Mirada Medical	LDM	⁹⁰ Y microspheres	CE/FDA
VelocityTM Varian	Varian	DPK/LD M	⁹⁰ Y microspheres	CE/FDA
RapidSphere v4.1				
Q-Suite v2.0	QUIREM Medical BV	LDM	¹⁶⁶ Ho microspheres	CE

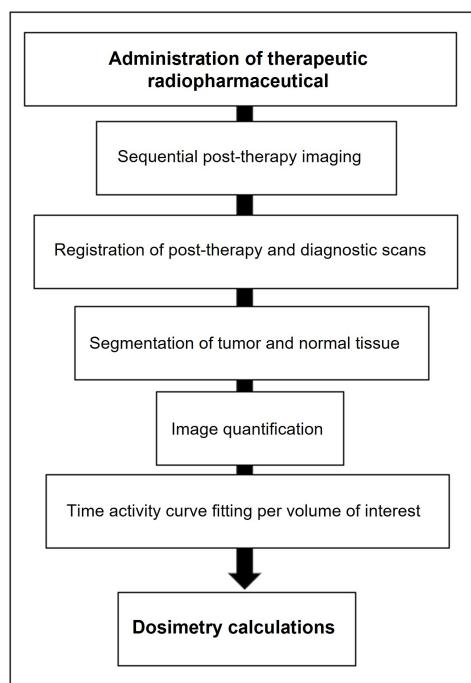


Figure 14. Schematic workflow for clinical dosimetry in RLT.

To overcome these issues, commercial software programs have been created to address challenges in phantom and patient-specific dosimetry (Tables 4 and 5). These programs offer advanced tools for precise radiation dose calculations and analysis, catering to medical physicists, radiation oncologists, and other healthcare professionals involved in radiation therapy planning. Phantom dosimetry involves measuring and computing radiation doses using phantoms that mimic human tissues and organs [107]. These phantoms validate RLT plans and delivery systems, providing features for phantom design, dose calculation, and analysis. Patient-specific dosimetry tailors RLT plans to individual patient characteristics, offering tools for precise dose calculation and optimization based on factors like patient anatomy and tumor location [103]. These programs use sophisticated algorithms to simulate radiation interaction with patient tissues, resulting in accurate dose distribution calculations.

The significance of ¹⁷⁷Lu-based SPECT/CT in internal dosimetry for NETs has surpassed that of

traditional diagnostic PET or SPECT imaging. This is mainly attributed to its direct connection to therapeutics within theranostics. Unlike standard dosimetry, which necessitates the identification of organs throughout the body and multiple sequential whole-body PET scans [108], this approach can pose challenges in routine theranostics practice due to the considerably longer half-lives of therapeutic radioisotopes such as ^{177}Lu compared to ^{18}F . A recent study has demonstrated that a dosimetry technique utilizing single-time point (STP) imaging exhibits reduced variability when compared to more intricate methods, such as employing multiple time points (MTP) for exponential fitting to determine residence time. Even though STP dosimetry is still under investigation, some studies have demonstrated that a dosimetry technique utilizing STP imaging could reduce variability when compared to more intricate methods, such as employing MTPs for exponential fitting to determine residence time. When dosimetry is performed with STP imaging instead of MTP imaging, a wider safety margin can be used for the kidney-absorbed dose limit. Protocols can also be designed to switch to MTP imaging in the next cycle as a cautionary measure only for those patients whose STP-estimated kidney-absorbed dose in the previous cycle is above a limit predetermined on the basis of the predicted error distribution for the specific STP model [103, 107, 109-111].

Given the strong correlation between internal dosimetry and the estimation of external radiation exposure, dosimetry plays a crucial role in Nuclear Medicine Physics (NMP) within the field of theranostics. Particularly, dosimetry holds significant promise in the investigation of ^{177}Lu -conjugated radiopharmaceuticals in clinical practice (Figure 14).

The emission of β -particles by ^{177}Lu serves to target and eliminate cancer cells, while the emission of γ -photons aids in imaging purposes. Research indicates that utilizing planar whole-body γ -camera protocols and SPECT/CT for multiple-time-point imaging of ^{177}Lu -DOTATATE enables quantitative evaluations of the radionuclide's distribution within individual patients over time. This approach facilitates personalized dosimetry calculations for both tumors and critical organs, such as the kidney. Theoretically, comprehending the absorbed dose could assist in adjusting the activity in subsequent treatment cycles appropriately and predicting outcomes. Furthermore, with the advancement of Deep Learning (DL) techniques, there exists the potential for automatically delineating target lesions and critical organs in this particular context.

Molecular imaging for dosimetry

The accumulation of a particular radiopharmaceutical is contingent upon the intensity and variability of target expression in both the tumor and normal organs, in addition to the biokinetic profile of the radiopharmaceutical [103]. Molecular imaging is essential for evaluating the biodistribution of radiopharmaceutical activity in individual patients undergoing treatment. Various imaging techniques, including SPECT, planar imaging, and hybrid imaging, can be utilized to assess the distribution of therapeutic radiopharmaceuticals. However, quantitative SPECT/CT is considered the optimal method for precise quantitative measurements [105]. The appropriate timing for imaging will be determined by the absorption and elimination patterns of the particular radiopharmaceutical agent. However, it is customary to acquire at least three SPECT time points following the administration of the treatment [104, 112]. These images serve the purpose of delineating specific areas of interest, where the organs at risk are accurately identified concerning the therapy being administered. By capturing a momentary snapshot of the concentration and distribution of the radiopharmaceutical, these images enable the creation of time-activity curves (TACs) for each region of interest (ROI). These curves provide a visual representation of how the activity accumulates, spreads, and is eliminated by a particular region throughout treatment. The area under the curve reflects the total accumulated activity, which, when multiplied by the energy released per decay and the absorbed fraction specific to the source and target, determines the absorbed dose. This absorbed dose per cumulated activity is known as an S factor, which represents a radiation transport factor unique to the source and target. In the case of normal organs, these factors are often computed through particle transport simulations and are commonly employed for estimating absorbed doses and assessing risks at a population level.

Voxel-based dosimetry

Organ-level dosimetry is crucial in the estimation of dose absorption, under the assumption of a uniform distribution of activity within organs and tumors. Conversely, sub-organ-level dosimetry, particularly at the voxel scale, takes into account patient-specific nonuniform activity distributions within organs and tumors. Voxel-wise dosimetry requires the calculation of energy deposition by transforming the three-dimensional activity distribution into a dose distribution through the utilization of data obtained from particle simulations (Figure 15).

This strategy represents the varied distribution of radiopharmaceuticals found in organs and tumors. Most methods for calculating voxel dose can be divided into four categories:

- The first and simplest way to calculate absorbed dose is that all doses are assumed to be: The associated radiation is absorbed locally, increasing the total decay number (TIAC).
- The second calculation method is based on the dose point kernel (DPK). TIAC images can be converted into dose distribution by collapsing the image in DPK. Another drawback is that DPK is dependent on voxel size. The DPK method cannot be considered as it is limited to one material.
- The third method is Monte Carlo simulation (MC).

- The fourth method is internal dosimetry using deep learning.

Voxel-based dosimetry technique was developed that takes into account the non-uniform activity distribution. This includes dose point kernels and voxel S-value (VSV) approaches. The dose point kernel represents radial absorbed dose in a homogeneous aqueous medium when there is an isotropic point source at that location. VSV is a voxel-level MIRD schema where sources and destinations are defined at the voxel level and then calculated in a 3D voxel matrix consisting of the aqueous medium. For more accurate individual dosimetry, voxel-based dosimetry is defined based on direct MC simulation. The MC simulation generates and tracks particles at the voxel level, calculates the stored energy, and absorbed dose at the voxel level (Figure 16).

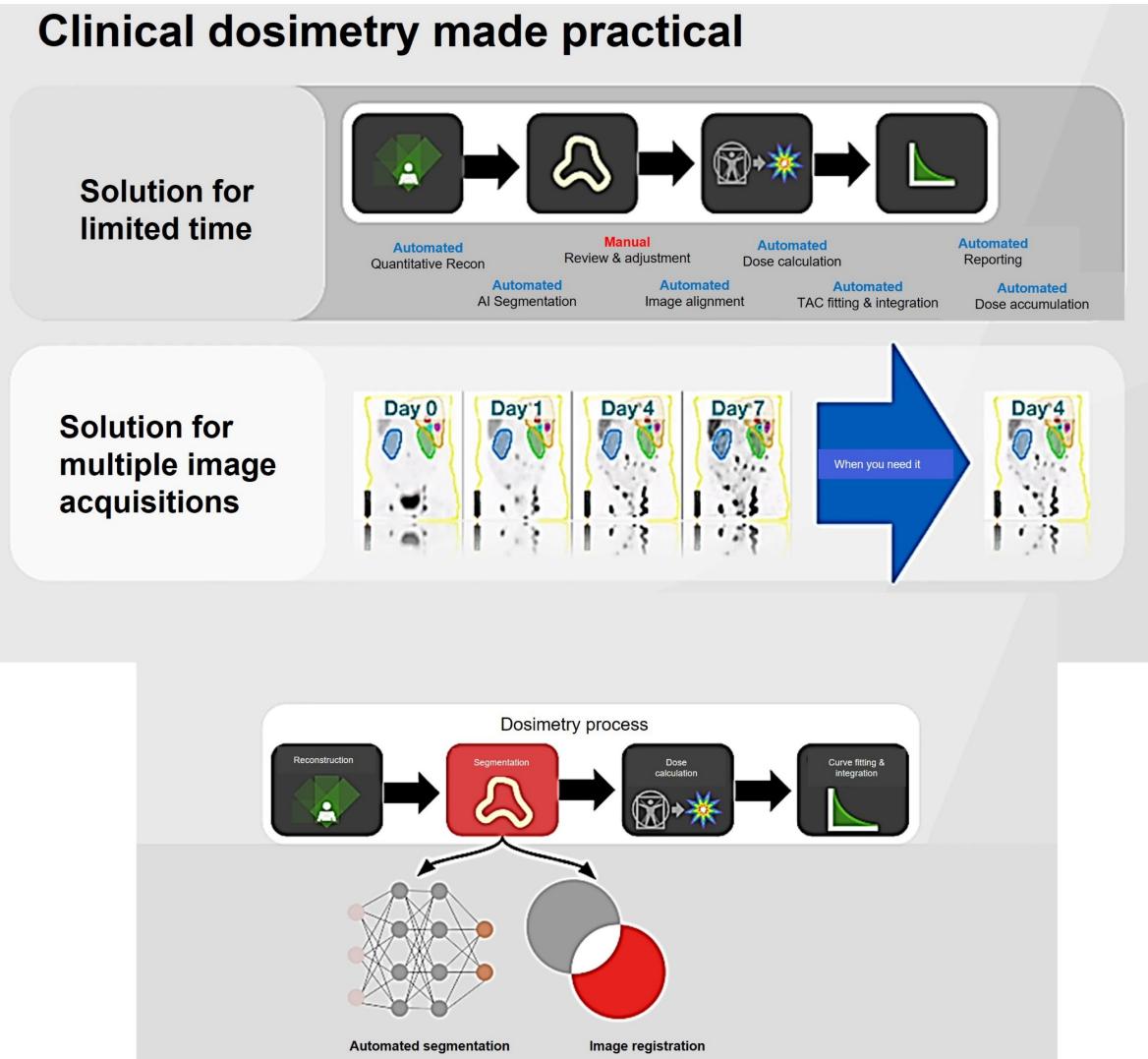


Figure 15. Voxel-wise dosimetry process (MiM dosimetry software's workflow).

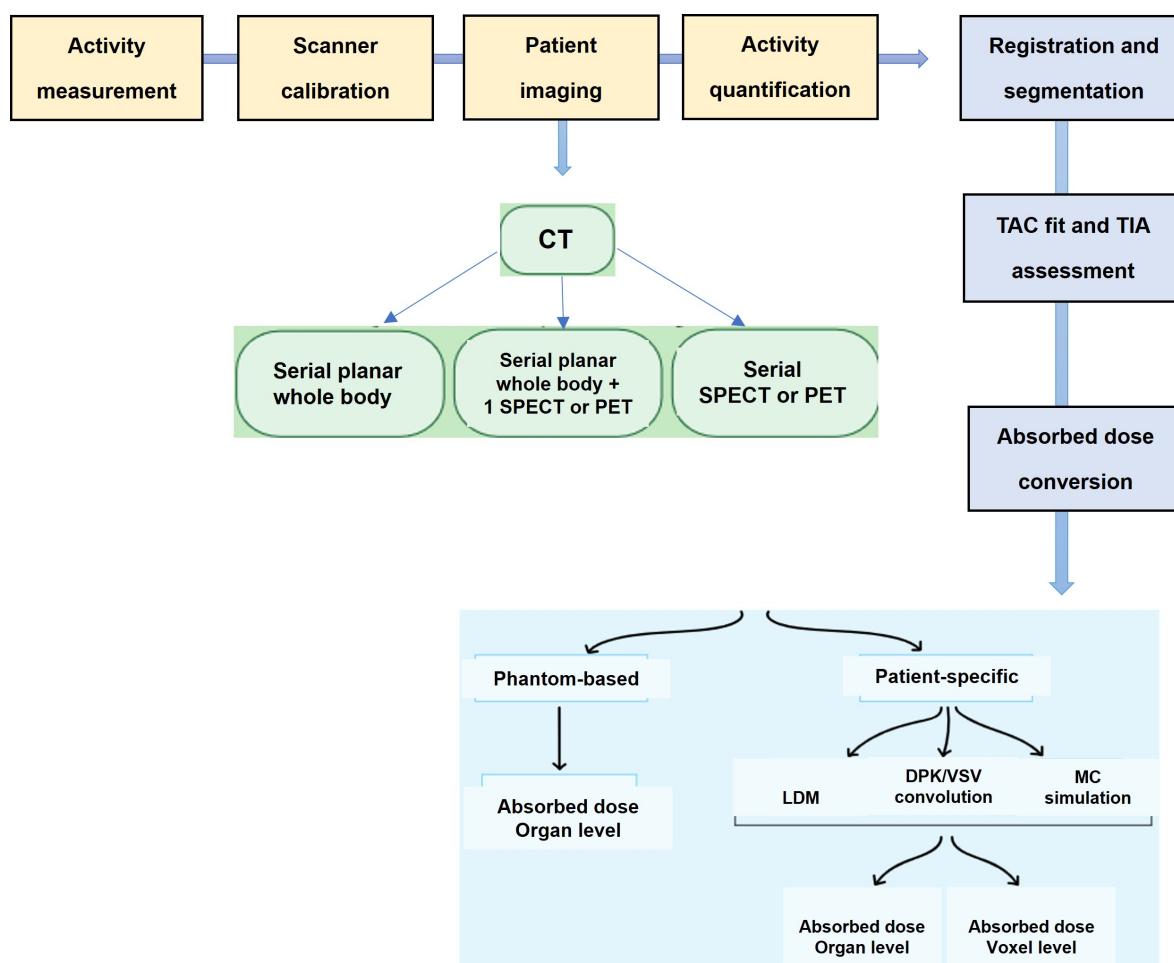


Figure 16. Schematic clinical dosimetric workflow.

The comparison between Ac-225 and Lu-177 dosimetry is essential due to the significant differences in their physical, chemical, and radiobiological properties, as well as the distinct challenges each presents. Ac-225, an alpha-emitter, offers high linear energy transfer (LET) radiation, which is highly effective for targeting micrometastases and single cancer cells. However, its complex decay chain, associated recoil effects, and redistribution of daughter isotopes pose substantial challenges for accurate dosimetry. On the other hand, Lu-177, a beta-emitter, provides more predictable energy deposition with established imaging capabilities using gamma emissions. These properties enable straightforward dosimetry using well-established macrodosimetric methods. Despite Lu-177's reliability in clinical applications, Ac-225 holds unique potential for treating specific cancer types where high LET radiation is beneficial. Hence, understanding the key differences and challenges in dosimetry between these radionuclides is crucial for optimizing therapeutic outcomes and advancing targeted radionuclide therapies.

a. Physical Properties

- Ac-225: An alpha-emitting radionuclide with high linear energy transfer (LET) radiation. Its decay chain produces multiple alpha particles and beta emissions, which contribute to its therapeutic efficacy but complicate dosimetry due to daughter redistribution and recoil effects [113].
- Lu-177: A beta-emitting radionuclide with low LET radiation, which allows for more predictable energy deposition over longer ranges. It emits gamma rays suitable for imaging, thus enabling more straightforward dosimetry [114].

b. Imaging and Quantification

- Ac-225: Alpha emissions are not directly imageable using standard nuclear imaging modalities. Indirect imaging relies on surrogate isotopes (e.g., Ac-227) or daughter radionuclides (e.g., Bi-213), which have different biodistribution profiles, leading to uncertainties in dosimetry [115].

- Lu-177: Gamma emissions at 113 keV and 208 keV are readily detected by SPECT, allowing accurate post-treatment imaging and quantification. Image-based dosimetry is well-established due to reliable gamma-ray detection [116].

c. Recoil Effects and Daughter Redistribution

- Ac-225: Alpha decay causes recoil of the parent nucleus, which can result in the release of daughter isotopes from the target tissue or carrier molecule, altering the dose distribution [115]. Redistribution of daughters like Bi-213 and Pb-211 into off-target organs, such as the kidneys, complicates dose estimates and increases toxicity risks [113].
- Lu-177: No significant recoil effects are observed, and the radionuclide remains localized, resulting in predictable dose distribution. Established radiopharmaceuticals, such as Lu-177-DOTATATE, have well-characterized biokinetics and dosimetry [117].

d. Dosimetry Methods

- Ac-225: Requires complex modeling due to the contribution of multiple alpha particles and beta emissions from daughter nuclides. Microdosimetry approaches are often used to estimate the absorbed dose at the cellular level due to high LET and short alpha-particle ranges [118].
- Lu-177: Dosimetry is based on well-established macrodosimetric methods using quantitative SPECT imaging and standard MIRD (Medical Internal Radiation Dose) models [119].
- Personalized dosimetry using voxel-based approaches is increasingly used for optimizing therapeutic efficacy [120].

e. Clinical Implications

- Ac-225: Suitable for targeting micrometastases and single cancer cells due to the localized high LET radiation. Dosimetry challenges limit widespread clinical application despite promising therapeutic outcomes [121].
- Lu-177: Widely used in targeted radionuclide therapy for neuroendocrine tumors and prostate cancer. Established dosimetry protocols facilitate clinical translation and regulatory approval [114].

$^{177}\text{Lu}/^{225}\text{Ac}$ -DOTATATE dosimetry

Currently, there is no established standard dosimetry for ^{177}Lu -DOTATATE in clinical practice.

Typically, patients receive a fixed-activity regimen of 7.4 GBq in four therapeutic infusions, regardless of how the radionuclide is distributed in their body, as long as they can tolerate the full dose. Although it is possible to perform ^{177}Lu -DOTATATE imaging during each therapy cycle, dosimetry can be utilized to estimate the optimal amount of activity to be injected in subsequent cycles. Despite observing a dose-response for ^{177}Lu -DOTATATE through dosimetry analysis, the NETTER-1 prospective trial, which is considered a clinically relevant trial, did not utilize or report dosimetry data to guide treatment decisions. Some smaller trials that did use dosimetry to guide therapy did not find a significant correlation between tumor response outcomes and the administered activity [122]. The upcoming COMPETE trial involving ^{177}Lu -DOTATOC (ClinicalTrials.gov no. NCT03049189) is set to utilize dosimetry data from a significant number of participants, potentially providing valuable insights into the practical application of this method. Furthermore, the forthcoming DOBATOC trial (ClinicalTrials.gov no. NCT04917484) aims to evaluate the effectiveness of dosimetry in determining treatment dosage as opposed to using standard fixed-dose regimens. With the increasing experience in somatostatin receptor-targeted radionuclide therapy, more intricate situations like prolonged or repeated treatment cycles have become apparent. A particular study employed dosimetry to extend the cycles of ^{177}Lu -DOTATATE until patients reached a specified renal dose threshold (23 Gy) or encountered other reasons for discontinuation, such as bone marrow toxicity or disease progression. Patients who tolerated higher doses demonstrated improved response rates and survival outcomes, though selection bias was a confounding factor [123]. Dosimetry holds great potential in the future for adjusting dosage regimens and predicting the patients who would benefit from additional therapy cycles. This aspect, which is currently lacking in the existing literature, remains to be explored in future trials of targeted radionuclide therapy for NETs and other types of cancer. In order to predict dosimetry for ^{225}Ac -DOTATATE, dosimetric data from ^{177}Lu -DOTATATE was utilized. The time-activity curves for ^{177}Lu -DOTATATE were adjusted to account for the physical half-life of ^{177}Lu and were then employed for predictive dosimetry for ^{225}Ac . To maintain equilibrium, residence times were estimated for all daughter isotopes in the ^{225}Ac chain. Furthermore, specific S-values were taken into consideration for each daughter isotope to accurately evaluate their contributions to the overall dose.

$^{177}\text{Lu}/^{225}\text{Ac-PSMA-617}$ dosimetry

In the same manner as the management of ^{177}Lu -DOTATATE, a fixed activity regimen is utilized for the administration of ^{177}Lu -PSMA-617 in current clinical practice. The implementation of dosimetry could provide significant prognostic information for radionuclide therapy in prostate cancer. Although dosimetry has been conducted for ^{177}Lu -PSMA-617 during each cycle in experimental settings, demonstrating the ability to predict PSA response, it has not been employed to direct dose administration for prostate cancer RLT and is not documented in prominent prospective trials such as TheraP or VISION [124, 125]. Ongoing endeavors are being made to establish uniform dosimetry techniques for radiopharmaceutical therapies. In this regard, collaborative guidelines have been developed by the EANM and the MIRD society, specifically focusing on the dosimetry of ^{177}Lu using quantitative SPECT. These guidelines have been successfully implemented in clinical trial settings [124]. Dosimetry is commonly required for the implementation of EBRT in clinical settings. Some nations are now recognizing the importance of dosimetry in conjunction with radiopharmaceuticals. Nevertheless, there is an ongoing discussion about whether dosimetry calculations should be mandatory in the clinical administration of ^{177}Lu -DOTATATE and ^{177}Lu -PSMA-617, as well as the advantages of tailoring therapeutic doses based on individual patient needs using dosimetry. A significant challenge in dosimetry for RLT is the limited ability to effectively image α -particle emitters. However, it is still possible to estimate dosimetry by utilizing a diagnostic surrogate radionuclide that emits γ rays, although it should be noted that changing the radionuclide could impact biodistribution [126]. These measurements have the potential to customize treatment by optimizing the correlation between dosage and response, all the while ensuring the safety of vital organs. It is of utmost importance to continue striving towards evaluating the influence of dosimetry on patient outcomes and establishing uniform techniques that can facilitate dosimetry in different medical facilities. These endeavors are essential to fully realize the potential advantages of dosimetry. The dosimetric data of ^{177}Lu -PSMA ligands were transformed into predictive dosimetry for ^{225}Ac -PSMA ligands, under the assumption of a comparable uptake pattern governed by the PSMA carrier, following the methodology previously described by Kratochwil *et al.* [121, 127]. The TACs for ^{177}Lu -PSMA were modified to accommodate the physical half-life of ^{177}Lu , and the resulting biological TACs were employed for the predictive dosimetry of ^{225}Ac . To

ensure equilibrium within the decay chain and assume no translocation during decay between subsequent disintegrations, the same residence time estimated for ^{225}Ac was applied to all the daughter isotopes in the ^{225}Ac chain. Additionally, specific S-values were taken into account for each daughter isotope to accurately evaluate their respective dose contributions. Drawing from existing literature, an RBE factor of 5 was utilized to assess the α -particle dose contribution in comparison to the γ and β emissions [128].

Whole-body voxel-based internal dosimetry using deep learning

Personalized medicine presents a new approach that seeks to enhance the effectiveness of healthcare and lower expenses. It holds great potential for tailoring diagnosis and treatment to each individual, thereby offering improved outcomes [129-132]. Precision medicine aims to shift from the prevailing one-size-fits-all approach to a personalized model. Within the realm of nuclear medicine, the calculation of dosage assumes a crucial role in aligning with this paradigm [107]. The precise determination of individualized dosages is crucial within this framework as it allows for the optimization of clinical protocols while reducing the risk of radiation-related side effects [133]. At present, the monitoring of patient dose in clinical settings frequently depends on simplified models, such as those obtained from the MIRD formalism [134]. The traditional MIRD method calculates organ-level dosimetry using time-integrated activity and radionuclide S-values to estimate the average absorbed dose per radioactive decay. This approach assumes consistent activity distribution within organs and ignores individual anatomical differences. To address variability in anatomy, newer techniques incorporate patient-specific computational models [135-138].

Furthermore, there have been advancements in voxel-based dosimetry methods, including the dose point kernel and VSV techniques [134]. Unlike probabilistic approaches, the dose point kernel method [139] computes the radial absorbed dose profile surrounding an isotropic point source within a uniform water medium [140, 141]. The MIRD schema at the voxel level is described as a 3D voxel matrix that shows the average absorbed dose to a target voxel for each unit of activity in a source voxel within an infinite homogeneous medium using MC simulations. Nevertheless, when performing voxel-based dose calculations, it is essential to take into account the non-uniform distribution of activity of the radiotracer, as well as the heterogeneity of the medium, which includes various material compositions such as lung,

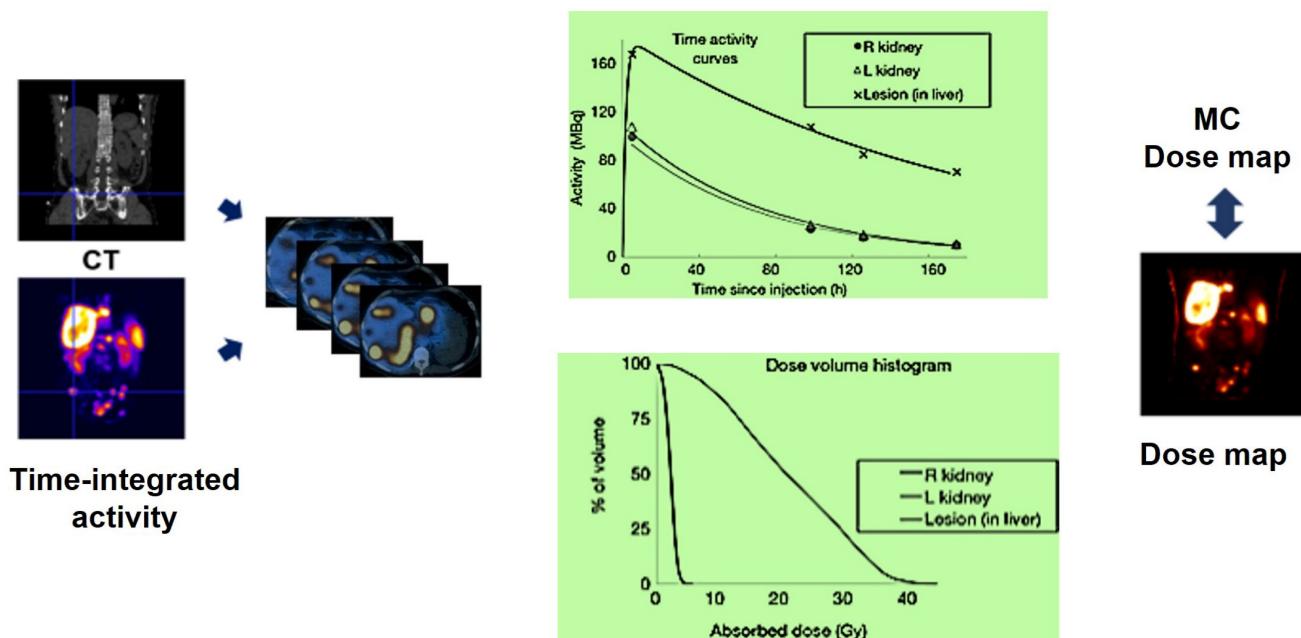


Figure 17. Sample dose volume histograms (DVHs) and lesion absorbed dose map corresponding to a patient imaged at 4 time points after cycle 1 of standard (7.4 GBq) ^{177}Lu -DOTATATE RLT. SPECT/CT images at each time point were input to a Monte Carlo dosimetry code and the corresponding dose-rate maps were integrated to derive the absorbed dose map [147].

soft tissue, and bone, that are often overlooked. To overcome this challenge, direct MC simulations, recognized as the standard for establishing a dependable dose calculation framework in a clinical environment, enable the precise calculation of a complete body dose map [142, 143].

MC simulation considers non-uniform activity distribution and patient-specific anatomical features, but it is limited by computational workload. Past research has explored MC simulations for personalized dosimetry in nuclear medicine [144-146]. The MC simulator uses hybrid PET/CT or SPECT/CT images to simulate radiation energy deposition from injected radiotracers, considering patient anatomy and voxelwise activity distribution (Figure 17).

Studies aim to balance voxel-scale dosimetry accuracy with computational efficiency [148, 149]. Khazaei Moghadam and colleagues proposed a method utilizing tissue-specific dose point kernels on a stylized phantom [150]. Building on this concept, Lee *et al.* extended the methodology by applying it to real patient data [151]. They incorporated various material densities into internal dose calculations, creating a range of voxelwise S-value kernels for different human body tissues. This method allowed for the generation of multiple voxel-scale dose maps similar to MIRD calculations. Each density-specific dose map was multiplied by the corresponding binary mask from CT-based segmentation to compute the final dose map. While this approach enhances dosimetry accuracy compared to single voxel S-value

methods, it assumes energy depositions primarily occur due to self-absorption, introducing potential errors at tissue boundaries. The importance of precise patient-specific dosimetry is highlighted by advancements in targeted radionuclide therapy and theranostic imaging [107]. MC simulation is considered the most accurate method for personalized dosimetry and is the gold standard for research, but its use in clinical settings is limited by its heavy computational requirements. Deep learning has emerged as a promising tool in computer vision and image processing, outperforming traditional methods in medical image analysis for PET and SPECT imaging. It excels in tasks such as attenuation and scatter correction [152-157], low-count image reconstruction [158-162], and automated image segmentation [138, 163-165].

Recently, deep learning techniques have been increasingly used in radiation dose estimation. Mardani *et al.* have introduced a new method using a multi-layer convolutional auto-encoder to predict dose distribution in external beam radiation therapy [166]. Nguyen *et al.* utilized a U-Net structure in their study on clinical treatment plan optimization, to improve the quality and consistency of treatment plans while also decreasing the computational time required [167]. Ma *et al.* successfully employed a deep learning approach to extract isodose characteristics in the context of modulated arc therapy treatment plans [168]. Kearney and colleagues introduced a three-dimensional fully convolutional algorithm

designed specifically for predicting doses in prostate stereotactic body radiotherapy patients [169]. Effective training of a deep learning algorithm relies on a well-defined ground truth as an essential ingredient [170]. The earlier influential studies used MC dosimetry as a substitute to establish ground truth for training the networks. However, this approach may result in inaccuracies due to simplifications in physical models [171]. Lee *et al.* employed a U-Net deep neural network architecture to address this constraint, utilizing training data derived from MC simulation for internal dosimetry purposes [172]. Gotz *et al.* used a modified U-Net network to estimate dose maps for patients given ¹⁷⁷Lu-PSMA ligands. They input CT images and static PET images into the network to predict a 3D dose map [173]. The training datasets in this research included a two-channel input: CT images for patient-specific density maps and MIRD-based voxel-scale dose maps from SPECT images. Ground truth was obtained from direct MC simulations. Previous studies used deep learning networks trained with whole-body dose maps from direct MC simulations, but creating a comprehensive training dataset was challenging due to the computational intensity of MC calculations. As a result, these studies either used a limited number of training samples or made approximations that could impact model accuracy.

Procedure guidelines or recommended practice

The establishment must have enough seating and essential amenities to ensure patient well-being. Chair use is typically limited to three infusions per day. We have expertise in chair breakdowns, radiation monitoring, and preparation procedures. Safety measures are customized to contain radioactivity and depend on the therapy and radionuclides used. For example, administering α -particle emitters like ²²³Ra dichloride is straightforward because α -particles travel a very short distance (<100 μ m) in most substances (Table 6) [174-176].

Therefore, minimal shielding and personal protective equipment are required for α -emitting RLT, allowing it to be administered in a clinic or office setting if desired [177, 178]. Conversely, β -emitting RLT, like ¹⁷⁷Lu vipivotide tetraxetan (Pluvicto) for prostate cancer, emits γ -photons in addition to β -particles, requiring more complex considerations for the facility [179, 180], especially in terms of potential daily exposure risks for medical personnel. A dedicated hot lab for storing and preparing Pluvicto doses is necessary, and specific therapy rooms must be designated. While some nuclear medicine centers

have lead-shielded private compartments for infusion chairs to enhance radiation protection, shielded patient rooms are not mandatory for Pluvicto administrations. However, patients receiving Pluvicto treatment should be isolated from others.

Patient scheduling

Patients scheduled for RLT need to be informed about potential treatment delays due to myelosuppression. Timely completion of necessary blood work is crucial to avoid cancellations and wasted doses. The complexity of maintaining timely administration is compounded by the decay profile of radioactive material and fixed treatment intervals. Supply chain interruptions for radiopharmaceuticals can also cause delays. Patients should be informed about possible schedule adjustments due to adverse events or supply chain disruptions. Providing logistical support can improve patient scheduling and access to RLT. The medical management plan for oncology therapies involves imaging, treatment, blood work, and follow-up appointments overseen by the prescribing clinician. Advanced Practice Providers can help manage RLT schedules and monitor patient progress. Collaboration between the oncology team and transfusion facility is essential for patients dependent on transfusions. Computerized workflow applications can optimize scheduling and prevent inefficiencies. Accurate forecasting is key to ensuring appropriate treatment [37, 179].

Imaging and RLT centers depend on a consistent supply of radiopharmaceutical agents, which are mainly produced in limited facilities like cyclotrons and nuclear reactors. Effective communication with suppliers is crucial due to the ordering timelines, with ¹⁷⁷Lu-based RLTs requiring orders to be placed at least two weeks in advance [37, 179]. Managing these timelines adds complexity to the supply chain, requiring additional personnel to promptly process orders through the Novartis Advanced Accelerator Applications Radiopharmaceutical Order Management Environment. This system helps minimize wastage, ensure timely patient care, and reduce the need for rescheduling. For Pluvicto, RLT centers coordinate their orders and supply based on a dosage regimen of 7.4 GBq administered every 6 weeks for up to 6 cycles via IV infusion [181].

Patient admission

Nurses play a crucial role in ensuring the smooth implementation of RLT treatments. Prior to the treatment day, nurses must confirm the schedule with the patient and engage in a comprehensive discussion regarding the planned procedures. This pre-treatment communication aims to educate and prepare the

patient, addressing any incontinence requirements (such as self-catheterization or the use of incontinence pads) to minimize potential radiation safety risks. Additionally, this conversation can also involve coordinating travel arrangements to the center, with certain centers assisting in accommodations the night before and/or after the treatment. The establishment of an efficient workflow is vital for the safety of both staff and patients, ultimately enhancing the patient's overall treatment experience (Figure 18). To ensure effective education, treatment, and supervision in RLT, dedicated staff should assist patients in getting acquainted with the therapy room, explaining safety protocols, and answering any questions.

Radioligand preparation

Prior to administration, a nuclear medicine technologist thoroughly examines the radioligand preparation to ensure the absence of any particulate matter. Subsequently, the technologist utilizes a dose calibrator to accurately measure the activity, guaranteeing that the patient receives the prescribed dose with precision. Following this, the radioligand is prepared for administration. If the radioligand is contained in a vial, it must be prepared in a hot lab before it can be transferred to a shielded syringe if the syringe method of administration is chosen [181]. In the case of Pluvicto, the standard approved activity of 7.4 GBq is typically administered to most patients [181]. However, this dosage can be adjusted if there are any risk factors that may pose a potential toxicity concern [127, 182].

Administration

Prior to administration, it is crucial to perform suitable imaging examinations, comprehensive blood counts, and assessments of kidney function. The

technique of RLT administration differs based on the particular RLT being employed. The equipment necessary for RLT administration, in adherence to radiation safety protocols, encompasses tongs, shielded barriers equipped with leaded glass, syringe shields, kits for handling radioactive material spills, and Geiger counters [179]. It is essential to have a treatment room with a patient restroom available at all times during the treatment period to accommodate frequent voiding needs. Patients who may experience incontinence should follow institutional guidelines treating excreta as radioactive waste. Moreover, all pre-medications and concomitant medications must be easily accessible. In the instance of Pluvicto, an antiemetic like oral ondansetron can be given 30–60 minutes before infusion and on days 2 and 3 of each treatment cycle [182]. Per the recommendations outlined by the EANM, corticosteroids may be administered starting one day prior to treatment and continued for multiple days post-treatment in individuals with metastases who are at risk of experiencing painful or obstructive swelling following RLT [182, 183]. In contrast to the use of amino acids for renal protection during Pluvicto treatment, which is necessary for neuroendocrine tumors treated with ¹⁷⁷Lu-DOTATATE, establishing proper venous access is crucial when administering RLT [181, 184]. The typical method of administering Pluvicto involves slowly pushing it through an IV catheter prefilled with 0.9% sterile sodium chloride solution over a period of approximately 1–10 minutes. Other methods of administration, such as the gravity method or the vial method, can also be used. To minimize the risk of extravasation, a saline flush of at least 10 mL is required before and after the infusion [184].

Table 6. Radiation safety requirements for radionuclides used in prostate cancer treatment.

Element	Tissue penetration (mm)	Radionuclide half-life	Specific material handling needs required to minimize radiation exposure
<i>α-emitting</i>			
²²³ Radium	< 0.1	11.4 d	Standard personal protection equipment, including mask, gloves, laboratory coat, safety glasses/goggles
²²⁵ Actinuma	0.05–0.08	10.0 d	
²²⁷ Thoriuma	0.05–0.08	18.7 d	
²¹¹ Astatinea	0.05	7.2 h	
²¹² Leada	0.06–0.09	10.6 h	
<i>β-emitting</i>			
¹³¹ Iodine	0.8	8.0 d	Shielding with an appropriate thickness of low atomic number ($Z < 14$) materials (e.g., Plexiglas) or aluminum
¹⁷⁷ Lutetium	0.67	6.6 d	Standard personal protection equipment, including gloves, laboratory coat, safety glasses/goggles
¹⁵³ Samariumb	0.40	46.5 h	
⁹⁰ Yttrium	5.30	64.1 h	
⁶⁷ Coppera	0.6	2.6 d	
¹⁶¹ Terbiuma	0.29	6.9 d	

Patients receiving Pluvicto treatment require close monitoring by a nurse under the supervision of the attending physician to address any issues efficiently. It means that immediate issues may arise during or after Pluvicto treatment, necessitating close monitoring. Pluvicto, being a radioligand therapy, can potentially cause acute side effects that require prompt management. These issues may include:

a. Infusion-Related Reactions: Patients might experience nausea, vomiting, or hypersensitivity reactions during or shortly after infusion.

b. Gastrointestinal Symptoms: Diarrhea or abdominal discomfort could occur, which may need immediate symptomatic treatment.

c. Radiation Safety Concerns: Handling potential radioactive contamination or addressing immediate radiation-related issues may also require attention.

Close monitoring ensures that any acute complications are promptly identified and managed, minimizing risks and improving patient safety during treatment.

Potential consequences may arise 3-4 weeks after therapy [184]. The most common side effects that are not immediate issues of Pluvicto are listed below [185]:

Hematological Toxicity: Pluvicto can lead to myelosuppression, manifesting as low blood counts (e.g., anemia, neutropenia, thrombocytopenia), which might require urgent intervention in severe cases. The initial two cycles of PSMA-RLT were generally well tolerated in terms of hematological effects. The degree of both qualitative and quantitative impairment of the bone marrow seems to correlate significantly with the burden of osseous tumors. Notably, only these patients with extensive bone involvement and a lack of therapeutic response experienced severe hematological adverse events, alongside a marked reduction in hematological parameters. This suggests that in individuals with mCRPC, a lack of response to PSMA-RLT may be a significant factor contributing to bone marrow impairment during the early stages of treatment.

Renal Function Impairment: As the kidneys are primarily responsible for excreting the radioactive component, monitoring for signs of acute kidney injury is crucial.

Common adverse reactions with an incidence of at least 20% include fatigue, dry mouth, nausea, anemia, decreased appetite, and constipation. Common laboratory abnormalities with an incidence of at least 30% include reductions in lymphocytes, hemoglobin, leukocytes, platelets, calcium, and sodium [181]. Adverse events, such as vomiting and diarrhea are frequently reported as well. Patients with extensive bone metastases may encounter bone marrow toxicity; however, it is important to note that

while a high osseous tumor burden is a risk factor, bone marrow toxicity can also occur in patients without bone metastases [184]. Cytopenia of clinical significance following Pluvicto treatment can be managed by administering transfusions, and the subsequent treatment cycle can be delayed until blood counts improve. Patients with bone metastasis that causes symptoms may experience a temporary increase in pain lasting for 3-7 days, which can be effectively managed with pain medication, steroids, and/or non-steroidal anti-inflammatory drugs. To monitor these symptoms and toxicities in prostate cancer patients undergoing Pluvicto treatment, nurses and physicians can utilize patient-reported outcome measures specifically designed for this type of therapy, such as the recently developed Functional Assessment of Cancer Therapy-Radionuclide Therapy (FACT-RNT) [186].

Patient discharge

Following the completion of treatment, the radiation safety officer (RSO) is responsible for assessing radiation levels at a distance of 1 meter from the patient. If the anticipated dose to any other individual is projected to be under $30 \mu\text{Sv}/\text{h}$, patients are permitted to travel long distances or proceed to the airport for their journey home, in accordance with our standard operating procedure. Discharge criteria for patients treated with ^{177}Lu -based therapies vary significantly across countries due to differences in regulatory frameworks and clinical practices. For example, in the United States, it is standard practice to discharge patients after ^{177}Lu -based therapy without requiring the Radiation Safety Officer (RSO) to measure radiation levels at a distance of 1 meter from the patient. Conversely, other countries may mandate stricter discharge protocols, including radiation monitoring, to comply with local radiation safety regulations. In an analysis by Demir *et al.*, the mean dose rate at 1 m, 4 hours, and 6 hours post-treatment for 23 patients receiving Pluvicto was $23 \pm 6 \mu\text{Sv}/\text{h}$ and $15 \pm 4 \mu\text{Sv}/\text{h}$, respectively [187]. Although ^{177}Lu emits relatively low levels of γ -radiation, the radiation protection guidelines for this isotope are not as strict as those for isotopes with higher γ -emission, such as iodine-131 [188]. Patients, caregivers, and healthcare professionals should receive detailed contamination control instructions during and after clinic visits. Patients must be aware of potential radioactive contamination from bodily fluids, as about half of Pluvicto activity is excreted in urine within four hours of treatment [189]. Patients should practice good bathroom hygiene and stay hydrated at home to achieve optimal clearance, given the typical duration of discharge [190]. Close contact with adults should be

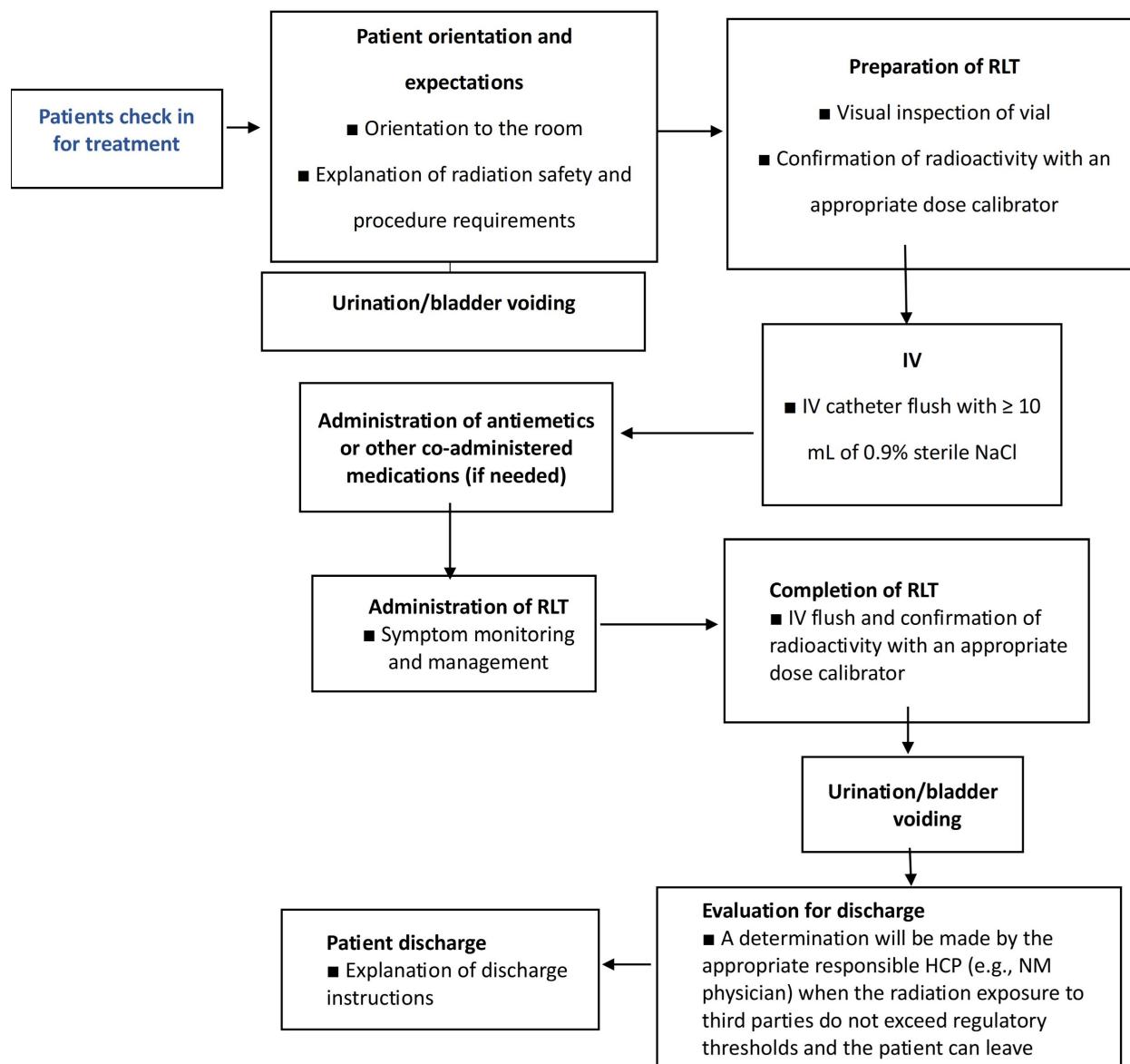


Figure 18. Patient's journey on treatment day. a. HCP, healthcare provider; IV, intravenous; NaCl, sodium chloride; NM, nuclear medicine; RLT, radioligand therapy. Advise patients to remain well hydrated and to urinate.

limited during the 48 hours following therapy. Contact with children and pregnant women should be avoided for at least 168 hours. These general guidelines aim to minimize radiation exposure while accounting for international variability in practices and ensuring compliance with radiation safety standards.

It is recommended to maintain separate sleeping arrangements from children for a week and from pregnant women for two weeks. Additionally, refraining from engaging in sexual activity is recommended for a duration of one week. Patients should promptly report any significant adverse events, especially myelosuppression symptoms, to the clinic and inform non-oncology healthcare providers about their Personalized Radiation Therapy.

Coordinated care inquiries should be directed to the oncology care team, as regular follow-up calls by the designated healthcare provider are crucial, given that some patients may not recognize severe adverse events.

Perspectives

As theranostics emerges as a pivotal approach in nuclear medicine and precision medicine, the establishment of standardized training programs becomes increasingly imperative [21]. These programs not only ensure safe and effective practice but also pave the way for future advancements. By fostering multidisciplinary collaboration among nuclear medicine professionals, qualified specialists, and healthcare experts, new technologies and

treatment modalities can be seamlessly integrated. Adhering to guiding principles will be essential for maintaining high standards, optimizing outcomes, and driving innovation [191]. Furthermore, specialized centers for radiotheranostic services are crucial, particularly in regions with limited access to innovative treatments. These centers play a vital role in bridging disparities between developed and developing nations, addressing challenges such as financial constraints and resource limitations [192]. Investing in professional training in dosimetry, radiochemistry, and radiopharmacy is key to enhancing treatment access, ensuring quality, and building specialized expertise and facilities in theranostics. The potential of combination theranostics with other targeted treatments may maximize synergistic benefits of personalized medicine but this requires careful examining through clinical trials [193].

Conclusion

The survival and quality of life for numerous patients are being significantly improved by the expanding landscape of radiotheranostics. This advancement has sparked the investigation and creation of new agents, as well as the broadening of indications for approved therapies. Although RPT represents a significant step towards precision medicine, there is still potential for further progress by gaining a deeper understanding of patient-specific factors that contribute to organ toxicity and tumor-related elements that influence treatment response. The customization of treatment for individual patients offers the potential to move closer to a paradigm of personalized disease management. In the current investigation, we present a thorough examination of the involvement of various radioisotopes in the induction of direct and indirect DNA damage, as well as their influence on the initiation of DNA repair mechanisms in cancer cells. Current data indicates that high-energy α -emitter radioisotopes have the potential to directly impact the DNA structure by causing ionization, resulting in the creation of ionized atoms or molecules. This ionization process predominantly leads to the formation of irreparable and intricate DSBs. On the other hand, the majority of DNA damage caused by β -emitting radioisotopes is indirect, as it involves the production of free radicals like ROS and subsequent chemical reactions. Beta particles themselves can also physically interact with the DNA molecule, resulting in SSBs and potentially reversible DSBs. Drawing on real-world experience, we offer practical guidance for the effective integration of RLT into current clinical practices. As the field of RLT continues to evolve

rapidly, robust RLT programs will need to adapt and refine the provided guidance in the future.

While this review provides a broad overview of multiple aspects of radiotheranostics, each section is supported by targeted references for readers who wish to explore specific topics in greater depth. The aim was that this structured approach facilitates both a comprehensive understanding and an easy transition to more detailed investigations in the field.

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Competing Interests

The authors have declared that no competing interest exists.

References

1. Walrand S, Hesse M, Lhommel R, Jamar F. The impact of image reconstruction bias on PET/CT 90Y dosimetry after radioembolization. *J Nucl Med*. 2015; 56: 494-5.
2. Muller C, Zhernosekov K, Koster U, Johnston K, Dorrer H, Hohn A, et al. A unique matched quadruplet of terbium radioisotopes for PET and SPECT and for alpha- and beta- radionuclide therapy: an *in vivo* proof-of-concept study with a new receptor-targeted folate derivative. *J Nucl Med*. 2012; 53: 1951-9.
3. Emani B, Lyman J, Brown A, Coia L, Goitein M, Munzenrider JE, et al. Tolerance of normal tissue to therapeutic irradiation. *Int J Radiat Oncol Biol Phys*. 1991; 21: 109-22.
4. Vergnaud L, Dewaraja YK, Giraudet AL, Badel JN, Sarrut D. A review of ^{177}Lu dosimetry workflows: how to reduce the imaging workloads? *EJNMMI Phys*. 2024; 11: 65.
5. Bagheri H, Mahdavi SR, Geramifar P, Neshasteh-Riz A, Sajadi Rad M, Dadgar H, et al. An update on the role of mpMRI and $(^{68}\text{Ga})\text{PSMA}$ PET imaging in primary and recurrent prostate cancer. *Clin Genitourin Cancer*. 2024; 22(3): 102076.
6. Maroufpor S, Aryana K, Nasseri S, Fazeli Z, Arabi H, Momennezhad M. Validation of dosimetry programs (Olinda & IDAC) for evaluation of absorbed dose in $(^{177}\text{Lu})\text{PSMA}$ therapy of metastatic castration-resistant prostate cancer (mCRPC) using Monte Carlo simulation. *EJNMMI Phys*. 2024; 11: 102.
7. Johnson KB, Wei WQ, Weeraratne D, Frisse ME, Misulis K, Rhee K, et al. Precision medicine, AI, and the future of personalized health care. *Clin Transl Sci*. 2021; 14: 86-93.
8. Arabi H, AkhavanAllaf A, Sanaat A, Shiri I, Zaidi H. The promise of artificial intelligence and deep learning in PET and SPECT imaging. *Phys Med*. 2021; 83: 122-37.
9. Zaidi H, El Naqa I. Quantitative molecular positron emission tomography imaging using advanced deep learning techniques. *Annu Rev Biomed Eng*. 2021; 23:249-76.
10. Gomes Marin JE, Nunes RF, Coutinho AM, Zaniboni EC, Costa LB, Barbosa FG, et al. Theranostics in nuclear medicine: emerging and re-emerging integrated imaging and therapies in the era of precision oncology. *Radiographics*. 2020; 40: 1715-40.
11. Ahn BC. Personalized medicine based on theranostic radioiodine molecular imaging for differentiated thyroid cancer. *Biomed Res Int*. 2016; 2016: 1680464.
12. Baum RP, Novruzov E, Zhao T, Greifenstein L, Jakobsson V, Perrone E, et al. Radiomolecular theranostics with fibroblast-activation-protein inhibitors and peptides. *Semin Nucl Med*. 2024; 54: 537-56.
13. Greifenstein L, Gunkel A, Hoehne A, Osterkamp F, Smerling C, Landvogt C, et al. 3BP-3940, a highly potent FAP-targeting peptide for theranostics - production, validation and first in human experience with Ga-68 and Lu-177. *iScience*. 2023; 26: 108541.
14. Choudhury P, Gupta M. Personalized & precision medicine in cancer: a theranostic approach. *Curr Radiopharm*. 2017; 10: 166-70.
15. Yordanova A, Eppard E, Kurpis S, Bundsuh RA, Schonberger S, Gonzalez-Carmona M, et al. Theranostics in nuclear medicine practice. *Oncotargets Ther*. 2017; 10: 4821-8.
16. Baum RP, Kulkarni HR, Carreras C. Peptides and receptors in image-guided therapy: theranostics for neuroendocrine neoplasms. *Semin Nucl Med*. 2012; 42: 190-207.
17. Baum RP, Kluge AW, Kulkarni H, Schorr-Neufing U, Niepsch K, Bitterlich N, et al. $[(^{177}\text{Lu})\text{DOTA}](0)\text{-D-Phe(1)-Tyr(3)-Octreotide}$ ($(^{177}\text{Lu})\text{DOTATOC}$) for

peptide receptor radiotherapy in patients with advanced neuroendocrine tumours: a phase-II study. *Theranostics*. 2016; 6: 501-10.

18. Zhang J, Kulkarni HR, Singh A, Schuchardt C, Niepsch K, Langbein T, et al. (177)Lu-PSMA-617 radioligand therapy in metastatic castration-resistant prostate cancer patients with a single functioning kidney. *J Nucl Med*. 2019; 60: 1579-86.
19. Kassis AI, Adelstein SJ. Radiobiologic principles in radionuclide therapy. *J Nucl Med*. 2005; 46 Suppl 1: 4-12.
20. Targeted Alpha Therapy Working Group, Parker C, Lewington V, Shore N, Kratochwil C, Levy M, et al. Targeted alpha therapy, an emerging class of cancer agents: a review. *JAMA Oncol*. 2018; 4: 1765-72.
21. Pascual TNB, Paez D, Iagaru A, Gnanasegaran G, Lee ST, Sathkege M, et al. Guiding principles on the education and practice of theranostics. *Eur J Nucl Med Mol Imaging*. 2024; 51: 2320-31.
22. Bushnell DL, Madsen MT, O'Cordisio T, Menda Y, Muzahir S, Ryan R, et al. Feasibility and advantage of adding (131)I-MIBG to (90)Y-DOTATOC for treatment of patients with advanced stage neuroendocrine tumors. *EJNMMI Res*. 2014; 4: 38.
23. Sgouros G, Bodei L, McDevitt MR, Nedrow JR. Radiopharmaceutical therapy in cancer: clinical advances and challenges. *Nat Rev Drug Discov*. 2020; 19: 589-608.
24. Ku A, Facca VJ, Cai Z, Reilly RM. Auger electrons for cancer therapy - a review. *EJNMMI Radiopharm Chem*. 2019; 4: 27.
25. Al-Ibraheem A, Scott AM. (161)Tb-PSMA unleashed: a promising new player in the theranostics of prostate cancer. *Nucl Med Mol Imaging*. 2023; 57: 168-71.
26. Mulford DA, Scheinberg DA, Jurcic JG. The promise of targeted alpha-particle therapy. *J Nucl Med*. 2005; 46 Suppl 1: 199-204.
27. Wahl RL, Sgouros G, Iravani A, Jacene H, Pryma D, Saboury B, et al. Normal-tissue tolerance to radiopharmaceutical therapies, the knowns and the unknowns. *J Nucl Med*. 2021; 62: 23-35.
28. Vallathol DH, Digumarthi R. An update on toxicity of therapeutic radionuclides. *Med Res Arch*. 2021; 9:4.
29. Schreuder N, de Romijn I, Jager PL, Kosterink JGW, van Puijenbroek EP. Safe use of radiopharmaceuticals in patients with chronic kidney disease: a systematic review. *EJNMMI Radiopharm Chem*. 2021; 6: 27.
30. Ladrière T, Faudemar J, Levigoureux E, Peyronnet D, Desmonts C, Vigne J. Safety and therapeutic optimization of lutetium-177 based radiopharmaceuticals. *Pharmaceutics*. 2023; 15: 1240.
31. Abbott A, Sakellis CG, Andersen E, Kuzuhara Y, Gilbert L, Boyle K, et al. Guidance on 177Lu-DOTATATE peptide receptor radionuclide therapy from the experience of a single nuclear medicine division. *J Nucl Med Technol*. 2018; 46: 237-44.
32. Hope TA, Antonarakis ES, Bodei L, Calais J, Iravani A, Jacene H, et al. SNMMI consensus statement on patient selection and appropriate use of 177Lu-PSMA-617 radionuclide therapy. *J Nucl Med*. 2023; 64: 1417-23.
33. Vlachostergios PJ, Niaz MJ, Skafida M, Mosallaei SA, Thomas C, Christos PJ, et al. Imaging expression of prostate-specific membrane antigen and response to PSMA-targeted beta-emitting radionuclide therapies in metastatic castration-resistant prostate cancer. *Prostate*. 2021; 81: 279-85.
34. Sharma R, Wang WM, Yusuf S, Evans J, Ramaswami R, Wernig F, et al. 68Ga-DOTATATE PET/CT parameters predict response to peptide receptor radionuclide therapy in neuroendocrine tumours. *Radiother Oncol*. 2019; 141: 108-15.
35. Seifert R, Emmett L, Rowe SP, Herrmann K, Hadaschik B, Calais J, et al. Second version of the prostate cancer molecular imaging standardized evaluation framework including response evaluation for clinical trials (PROMISE V2). *Eur Urol*. 2023; 83: 405-12.
36. Sartor O, de Bono J, Chi KN, Fizazi K, Herrmann K, Rahbar K, et al. Lutetium-177-PSMA-617 for metastatic castration-resistant prostate cancer. *N Engl J Med*. 2021; 385: 1091-103.
37. Burkett BJ, Dundar A, Young JR, Packard AT, Johnson GB, Halfdanarson TR, et al. How we do it: a multidisciplinary approach to 177Lu DOTATATE peptide receptor radionuclide therapy. *Radiology*. 2021; 298: 261-74.
38. Strosberg J, El-Haddad G, Wolin E, Hendifar A, Yao J, Chasen B, et al. Phase 3 trial of 177Lu-Dotatate for midgut neuroendocrine tumors. *N Engl J Med*. 2017; 376: 125-35.
39. Strosberg JR, Caplin ME, Kunz PL, Ruszniewski PB, Bodei L, Hendifar A, et al. 177Lu-Dotatate plus long-acting octreotide versus high-dose long-acting octreotide in patients with midgut neuroendocrine tumours (NETTER-1): final overall survival and long-term safety results from an open-label, randomised, controlled, phase 3 trial. *Lancet Oncol*. 2021; 22: 1752-63.
40. Yadav MP, Ballal S, Sahoo RK, Bal C. Efficacy and safety of (225)Ac-DOTATATE targeted alpha therapy in metastatic paragangliomas: a pilot study. *Eur J Nucl Med Mol Imaging*. 2022; 49: 1595-606.
41. Kanellopoulos P, Mattsson A, Abouzayed A, Obeid K, Nock BA, Tolmachev V, et al. Preclinical evaluation of new GRPR-antagonists with improved metabolic stability for radiotherapeutic use in oncology. *EJNMMI Radiopharm Chem*. 2024; 9: 13.
42. Echavide W, Fagret D, Faraggi M, Picco V, Montemagno C. Recent pre-clinical advancements in nuclear medicine: pioneering the path to a limitless future. *Cancers (Basel)*. 2023; 15: 4839.
43. Hofman MS, Emmett L, Violet J, Y. Zhang A, Lawrence NJ, Stockler M, et al. TheraP: a randomized phase 2 trial of 177Lu-PSMA-617 theranostic treatment vs cabazitaxel in progressive metastatic castration-resistant prostate cancer (clinical trial protocol ANZUP 1603). *BJU Int*. 2019; 124: 5-13.
44. Emmett L, Willowson K, Violet J, Shin J, Blanksby A, Lee J. Lutetium (177)PSMA radionuclide therapy for men with prostate cancer: a review of the current literature and discussion of practical aspects of therapy. *J Med Radiat Sci*. 2017; 64: 52-60.
45. Mullard A. FDA approves first PSMA-targeted radiopharmaceutical. *Nat Rev Drug Discov*. 2022; 21: 327.
46. Kratochwil C, Brucherseifer F, Giesel FL, Weis M, Verburg FA, Mottaghay F, et al. 225Ac-PSMA-617 for PSMA-targeted alpha-radiation therapy of metastatic castration-resistant prostate cancer. *J Nucl Med*. 2016; 57: 1941-4.
47. Sapathe S, Sood A, Das CK, Mittal BR. Evolving role of 225Ac-PSMA radioligand therapy in metastatic castration-resistant prostate cancer—a systematic review and meta-analysis. *Prostate Cancer Prostatic Dis*. 2021; 24: 880-90.
48. Lepareur N, Ramee B, Mougin-Degraef M, Bourgeois M. Clinical advances and perspectives in targeted radionuclide therapy. *Pharmaceutics*. 2023; 15: 1733.
49. Baatout S. Radiobiology textbook. Cham, Switzerland: Springer International Publishing; 2023 Sep 23.
50. Sminia P, Guipaud O, Viktorsson K, Ahire V, Baatout S, Boterberg T, et al. Clinical radiobiology for radiation oncology. *Radiobiology Textbook*. Cham: Springer International Publishing; 2023. 237-309.
51. Adamo A, Collis SJ, Adelman CA, Silva N, Horejsi Z, Ward JD, et al. Preventing nonhomologous end joining suppresses DNA repair defects of Fanconi anemia. *Mol cell*. 2010; 39: 25-35.
52. Yousefzadeh M, Henpita C, Vyas R, Soto-Palma C, Robbins P, Niedernhofer L. DNA damage-how and why we age? *eLife*. 2021; 10: 62852.
53. Groelly FJ, Fawkes M, Dagg RA, Blackford AN, Tarsounas M. Targeting DNA damage response pathways in cancer. *Nat Rev Cancer*. 2023; 23: 78-94.
54. Tubbs A, Nussenzeig A. Endogenous DNA damage as a source of genomic instability in cancer. *Cell*. 2017; 168: 644-56.
55. De Boni R, van Larebeks N. Endogenous DNA damage in humans: a review of quantitative data. *Mutagenesis*. 2004; 19: 169-85.
56. Roos WP, Thomas AD, Kaina B. DNA damage and the balance between survival and death in cancer biology. *Nat Rev Cancer*. 2016; 16: 20-33.
57. Antonangeli F, Natalini A, Garassino M, C, Sica A, Santoni A, Di Rosa F. Regulation of PD-L1 expression by NF- κ B in cancer. *Front Immunol*. 2020; 11: 584626.
58. Alhmoud JF, Woolley JF, Al Moustafa AE, Malki MI. DNA damage/repair management in cancers. *Cancers (Basel)*. 2020; 12: 1050.
59. Santivasi WL, Xia F. Ionizing radiation-induced DNA damage, response, and repair. *Antioxid Redox Signal*. 2014; 21: 251-9.
60. Curtin NJ. Inhibiting the DNA damage response as a therapeutic manoeuvre in cancer. *Br J Pharmacol*. 2013; 169: 1745-65.
61. Pearl LH, Schierz AC, Ward SE, Al-Lazikani B, Pearl FM. Therapeutic opportunities within the DNA damage response. *Nat Rev Cancer*. 2015; 15: 166-80.
62. Ziegler JF. Comments on ICRU report no. 49: stopping powers and ranges for protons and alpha particles. *Radiat Res*. 1999; 152(2):219-22.
63. Kassis AI, Harris CR, Adelstein SJ, Ruth TJ, Lambrecht R, Wolf AP. The in vitro radiobiology of astatin-211 decay. *Radiat Res*. 1986; 105: 27-36.
64. Aghamohammadi SZ, Goodhead DT, Savage JR. Induction of sister chromatid exchanges (SCE) in G0 lymphocytes by plutonium-238 alpha-particles. *Int J Radiat Biol Relat Stud Phys Chem Med*. 1988; 53: 909-15.
65. Goodhead DT. Molecular and cell models of biological effects of heavy ion radiation. *Radiat Environ Biophys*. 1995; 34: 67-72.
66. Gudkov SV, Shlyagina NY, Vodeneev VA, Zvyagin AV. Targeted radionuclide therapy of human tumors. *Int J Mol Sci*. 2015; 17: 33.
67. Bishayee A, Rao DV, Bouchet LG, Bolch WE, Howell RW. Protection by DMSO against cell death caused by intracellularly localized iodine-125, iodine-131 and polonium-210. *Radiat Res*. 2000; 153: 416-27.
68. Asadian S, Mirzaei H, Kalantari BA, Davarpanah MR, Mohamadi M, Shpichka A, et al. Beta-radiating radionuclides in cancer treatment, novel insight into promising approach. *Pharmacol Res*. 2020; 160: 105070.
69. Filippi L, Schillaci O, Cianni R, Bagni O. Yttrium-90 resin microspheres and their use in the treatment of intrahepatic cholangiocarcinoma. *Future Oncol*. 2018; 14: 809-18.
70. Schumann S, Scherthan H, Lapa C, Serfling S, Muhtadi R, Lassmann M, et al. DNA damage in blood leucocytes of prostate cancer patients during therapy with (177)Lu-PSMA. *Eur J Nucl Med Mol Imaging*. 2019; 46: 1723-32.
71. Eberlein U, Nowak C, Bluemel C, Buck AK, Werner RA, Scherthan H, et al. DNA damage in blood lymphocytes in patients after (177)Lu peptide receptor radionuclide therapy. *Eur J Nucl Med Mol Imaging*. 2015; 42: 1739-49.
72. Reynolds TS, Bandari RP, Jiang Z, Smith CJ. Lutetium-177 labeled bombesin peptides for radionuclide therapy. *Curr Radiopharm*. 2016; 9: 33-43.
73. Panigone S, Nunn AD. Lutetium-177-labeled gastrin releasing peptide receptor binding analogs: a novel approach to radionuclide therapy. *Q J Nucl Med Mol Imaging*. 2006; 50: 310-21.
74. Purohit NK, Shah RG, Adant S, Hoepfner M, Shah GM, Beauregard JM. Potentiation of (177)Lu-octreotide peptide receptor radionuclide therapy of human neuroendocrine tumor cells by PARP inhibitor. *Oncotarget*. 2018; 9: 24693-706.
75. Khanha A. DNA damage in cancer therapeutics: a boon or a curse? *Cancer Res*. 2015; 75: 2133-8.
76. Chinol M, Vallabhanjosula S, Goldsmith SJ, Klein MJ, Deutsch KF, Chinen LK, et al. Chemistry and biological behavior of samarium-153 and

rhenium-186-labeled hydroxyapatite particles: potential radiopharmaceuticals for radiation synovectomy. *J Nucl Med.* 1993; 34: 1536-42.

77. Wang C, Wang J, Jiang H, Zhu M, Chen B, Bao W. In vitro study on apoptosis induced by strontium-89 in human breast carcinoma cell line. *J Biomed Biotechnol.* 2011; 2011: 541487.
78. Paschalais A, Sheehan B, Riisnaes R, Rodrigues DN, Gurel B, Bertan C, et al. Prostate-specific membrane antigen heterogeneity and DNA repair defects in prostate cancer. *Eur Urol.* 2019; 76: 469-78.
79. van der Dolen MJ, Mehra N, van Oort IM, Looijen-Salamon MG, Janssen MJR, Custers JAE, et al. Clinical outcomes and molecular profiling of advanced metastatic castration-resistant prostate cancer patients treated with (225)Ac-PSMA-617 targeted alpha-radiation therapy. *Urol Oncol.* 2021; 39: 729.
80. Sancar A, Lindsey-Boltz LA, Unsal-Kacmaz K, Linn S. Molecular mechanisms of mammalian DNA repair and the DNA damage checkpoints. *Annu Rev Biochem.* 2004; 73: 39-85.
81. Shrivastav M, De Haro LP, Nickoloff JA. Regulation of DNA double-strand break repair pathway choice. *Cell Res.* 2008; 18: 134-47.
82. Haro KJ, Scott AC, Scheinberg DA. Mechanisms of resistance to high and low linear energy transfer radiation in myeloid leukemia cells. *Blood.* 2012; 120: 2087-97.
83. Yard BD, Gopal P, Bannik K, Siemeister G, Hagemann UB, Abazeed ME. Cellular and genetic determinants of the sensitivity of cancer to alpha-particle irradiation. *Cancer Res.* 2019; 79: 5640-51.
84. King AP, Lin FL, Escoria FE. Why bother with alpha particles? *Eur J Nucl Med Mol Imaging.* 2021; 49: 7-17.
85. Pouget JP, Lozza C, Deshayes E, Boudousq V, Navarro-Teulon I. Introduction to radiobiology of targeted radionuclide therapy. *Front Med.* 2015; 2: 12.
86. Solanki JH, Tritt T, Pasternack JB, Kim JJ, Leung CN, Domogauer JD, et al. Cellular response to exponentially increasing and decreasing dose rates: implications for treatment planning in targeted radionuclide therapy. *Radiat Res.* 2017; 188: 221-34.
87. Neumaier T, Swenson J, Pham C, Polyzos A, Lo AT, Yang P, et al. Evidence for formation of DNA repair centers and dose-response nonlinearity in human cells. *Proc Natl Acad Sci U S A.* 2012; 109: 443-8.
88. Rothkamm K, Lobrich M. Evidence for a lack of DNA double-strand break repair in human cells exposed to very low x-ray doses. *Proc Natl Acad Sci U S A.* 2003; 100: 5057-62.
89. Grudzenki S, Rathis A, Conrad S, Rube CE, Lobrich M. Inducible response required for repair of low-dose radiation damage in human fibroblasts. *Proc Natl Acad Sci U S A.* 2010; 107: 14205-10.
90. Rothkamm K, Barnard S, Moquet J, Ellender M, Rana Z, Burdak-Rothkamm S. DNA damage foci: Meaning and significance. *Environ Mol Mutagen.* 2015; 56: 491-504.
91. Tubiana M, Aurengo A, Averbeck D, Masse R. The debate on the use of linear no threshold for assessing the effects of low doses. *J Radiol Prot.* 2006; 26: 317-24.
92. Lobrich M, Rief N, Kuhne M, Heckmann M, Fleckenstein J, Rube C, et al. In vivo formation and repair of DNA double-strand breaks after computed tomography examinations. *Proc Natl Acad Sci U S A.* 2005; 102: 8984-9.
93. Brooks AL, Hoel DG, Preston RJ. The role of dose rate in radiation cancer risk: evaluating the effect of dose rate at the molecular, cellular and tissue levels using key events in critical pathways following exposure to low LET radiation. *Int J Radiat Biol.* 2016; 92: 405-26.
94. Blyth BJ, Sykes PJ. Radiation-induced bystander effects: what are they, and how relevant are they to human radiation exposures? *Radiat Res.* 2011; 176: 139-57.
95. Nagasawa H, Little JB. Induction of sister chromatid exchanges by extremely low doses of alpha-particles. *Cancer Res.* 1992; 52: 6394-6.
96. Brady D, O'Sullivan JM, Prise KM. What is the role of the bystander response in radionuclide therapies? *Front Oncol.* 2013; 3: 215.
97. Widel M. Radionuclides in radiation-induced bystander effect; may it share in radionuclide therapy? *Neoplasma.* 2017; 64: 641-54.
98. Sundlov A, Sjogren-Gleisner K, Svensson J, Ljungberg M, Olsson T, Bernhardt P, et al. Individualised (177)Lu-DOTATATE treatment of neuroendocrine tumours based on kidney dosimetry. *Eur J Nucl Med Mol Imaging.* 2017; 44: 1480-9.
99. Chiesa C, Sjogren Gleisner K, Flux G, Gear J, Walrand S, Bacher K, et al. The conflict between treatment optimization and registration of radiopharmaceuticals with fixed activity posology in oncological nuclear medicine therapy. *Eur J Nucl Med Mol Imaging.* 2017; 44: 1783-6.
100. Frey EC, Humm JL, Ljungberg M. Accuracy and precision of radioactivity quantification in nuclear medicine images. *Semin Nucl Med.* 2012; 42: 208-18.
101. Marin G, Vanderlinde B, Karfis I, Guiot T, Wimana Z, Flamen P, et al. Accuracy and precision assessment for activity quantification in individualized dosimetry of (177)Lu-DOTATATE therapy. *EJNMMI Phys.* 2017; 4: 7.
102. Hippelainen E, Tenhunen M, Maenpaa H, Sohlberg A. Quantitative accuracy of (177)Lu SPECT reconstruction using different compensation methods: phantom and patient studies. *EJNMMI Res.* 2016; 6: 16.
103. Lassmann M, Eberlein U. The relevance of dosimetry in precision medicine. *J Nucl Med.* 2018; 59: 1494-9.
104. Dewaraja YK, Frey EC, Sgouros G, Brill AB, Roberson P, Zanzonico PB, et al. MIRD pamphlet No. 23: quantitative SPECT for patient-specific 3-dimensional dosimetry in internal radionuclide therapy. *J Nucl Med.* 2012; 53: 1310-25.
105. Ljungberg M, Celler A, Konijnenberg MW, Eckerman KF, Dewaraja YK, Sjogren-Gleisner K, et al. MIRD Pamphlet No. 26: Joint EANM/MIRD guidelines for quantitative 177Lu SPECT applied for dosimetry of radiopharmaceutical therapy. *J Nucl Med.* 2016; 57: 151-62.
106. Huizing DMV, de Wit-van der Veen BJ, Verheij M, Stokkel MPM. Dosimetry methods and clinical applications in peptide receptor radionuclide therapy for neuroendocrine tumours: a literature review. *EJNMMI Res.* 2018; 8: 89.
107. Stabin MG, Madsen MT, Zaidi H. Personalized dosimetry is a must for appropriate molecular radiotherapy. *Med Phys.* 2019; 46: 4713-6.
108. Kesavan M, Grover P, Lam WS, Claringbold PG, Turner JH. Long-term hematologic toxicity of 177Lu-octreotate-capecitabine-temozolomide therapy of GEPNET. *Endocr Relat Cancer.* 2021; 28: 521-7.
109. Shi M, Jakobsson V, Greifenstein L, Khong P, Chen X, Baum R. Alpha-peptide receptor radionuclide therapy using actinium-225 labeled somatostatin receptor agonists and antagonists. *Front Med.* 2022; 9: 1034315.
110. Hardiansyah D, Riana A, Beer AJ, Glattling G. Single-time-point estimation of absorbed doses in PRRT using a non-linear mixed-effects model. *Z Med Phys.* 2023; 33: 70-81.
111. Brosch-Lenz J, Delker A, Volter F, Unterrainer LM, Kaiser L, Bartenstein P, et al. Toward single-time-point image-based dosimetry of (177)Lu-PSMA-617 therapy. *J Nucl Med.* 2023; 64: 767-74.
112. Siegel JA, Thomas SR, Stubbs JB, Stabin MG, Hays MT, Koral KF, et al. MIRD pamphlet no. 16: Techniques for quantitative radiopharmaceutical biodistribution data acquisition and analysis for use in human radiation dose estimates. *J Nucl Med.* 1999; 40: 37-61.
113. Kratochwil C, Bruchertseifer F, Giesel FL, Weis M, Verburg FA, Mottaghay F, et al. 225Ac-PSMA-617 for PSMA-targeted α -radiation therapy of metastatic castration-resistant prostate cancer. *J Nucl Med.* 2016; 57: 1941-4.
114. Fendler WP, Stupar AD, Evans-Axelson S, Luckeath K, Wei L, Kim W, et al. Establishing (177)Lu-PSMA-617 radioligand therapy in a syngeneic model of murine prostate cancer. *J Nucl Med.* 2017; 58: 1786-92.
115. Hassfjell S, Brechbiel MW. The development of the alpha-particle emitting radionuclides 212Bi and 213Bi, and their decay chain related radionuclides, for therapeutic applications. *Chem Rev.* 2001; 101: 2019-36.
116. Delker A, Schleske M, Liubchenko G, Berg I, Zacherl MJ, Brendel M, et al. Biodistribution and dosimetry for combined [(177)Lu]Lu-PSMA-I&T/[(225)Ac]Ac-PSMA-I&T therapy using multi-isotope quantitative SPECT imaging. *Eur J Nucl Med Mol Imaging.* 2023; 50: 1280-90.
117. Sgouros G, Dewaraja YK, Escoria F, Graves SA, Hope TA, Iravani A, et al. Tumor response to radiopharmaceutical therapies: the knowns and the unknowns. *J Nucl Med.* 2021; 62: 125-225.
118. Hindorf C, Emfietzoglou D, Linden O, Bousis C, Fotopoulos A, Kostarelos K, et al. Single-cell dosimetry for radioimmunotherapy of B-cell lymphoma patients with special reference to leukemic spread. *Cancer Biother Radiopharm.* 2007; 22: 357-66.
119. Stabin MG, Xu XG, Emmons MA, Segars WP, Shi C, Fernald MJ. RADAR reference adult, pediatric, and pregnant female phantom series for internal and external dosimetry. *J Nucl Med.* 2012; 53: 1807-13.
120. Sandstrom M, Garske-Roman U, Granberg D, Johansson S, Widstrom C, Eriksson B, et al. Individualized dosimetry of kidney and bone marrow in patients undergoing 177Lu-DOTA-octreotate treatment. *J Nucl Med.* 2013; 54: 33-41.
121. Kratochwil C, Schmidt K, Afshar-Oromieh A, Bruchertseifer F, Rathke H, Morgenstern A, et al. Targeted alpha therapy of mCRPC: dosimetry estimate of (213)Bismuth-PSMA-617. *Eur J Nucl Med Mol Imaging.* 2018; 45: 31-7.
122. Del Prete M, Buteau FA, Beauregard JM. Personalized (177)Lu-octreotate peptide receptor radionuclide therapy of neuroendocrine tumours: a simulation study. *Eur J Nucl Med Mol Imaging.* 2017; 44: 1490-500.
123. Garske-Roman U, Sandstrom M, Fross Baron K, Lundin L, Hellman P, Welin S, et al. Prospective observational study of (177)Lu-DOTA-octreotate therapy in 200 patients with advanced metastasized neuroendocrine tumours (NETs): feasibility and impact of a dosimetry-guided study protocol on outcome and toxicity. *Eur J Nucl Med Mol Imaging.* 2018; 45: 970-88.
124. Chen Z, Nath R. Biologically effective dose (BED) for interstitial seed implants containing a mixture of radionuclides with different half-lives. *Int J Radiat Oncol Biol Phys.* 2003; 55: 825-34.
125. Dahle J, Bruland OS, Larsen RH. Relative biologic effects of low-dose-rate alpha-emitting 227Th-rituximab and beta-emitting 90Y-tixutxetan-irbitumomab versus external beam X-radiation. *Int J Radiat Oncol Biol Phys.* 2008; 72: 186-92.
126. Sarnelli A, Belli ML, Di Iorio V, Mezzenga E, Celli M, Severi S, et al. Dosimetry of (177)Lu-PSMA-617 after mannitol infusion and glutamate tablet administration: preliminary results of EUDRACT/RSO 2016-002732-32 IRST protocol. *Molecules.* 2019; 24: 621.
127. Kratochwil C, Bruchertseifer F, Rathke H, Bronzel M, Apostolidis C, Weichert W, et al. Targeted alpha-therapy of metastatic castration-resistant prostate cancer with (225)Ac-PSMA-617: dosimetry estimate and empiric dose finding. *J Nucl Med.* 2017; 58: 1624-31.
128. Sgouros G, Roeske JC, McDevitt MR, Palm S, Allen BJ, Fisher DR, et al. MIRD Pamphlet No. 22 (abridged): radiobiology and dosimetry of alpha-particle emitters for targeted radionuclide therapy. *J Nucl Med.* 2010; 51: 311-28.
129. Baumann M, Krause M, Overgaard J, Debus J, Bentzen SM, Daartz J, et al. Radiation oncology in the era of precision medicine. *Nat Rev Cancer.* 2016; 16: 234-49.

130. Akhavanallaf A, Shiri I, Arabi H, Zaidi H. Whole-body voxel-based internal dosimetry using deep learning. *Eur J Nucl Med Mol Imaging*. 2021; 48: 670-82.

131. Khoshyari-morad Z, Jahangir R, Miri-Hakimabad H, Mohammadi N, Arabi H. Monte carlo-based estimation of patient absorbed dose in 99mTc-DMSA-, MAG3, and-DTPA SPECT imaging using the university of Florida (UF) phantoms. *Appl Radiat Isot*. 2025; 220: 111772.

132. Akhavanallaf A, Mohammadi R, Shiri I, Salimi Y, Arabi H, Zaidi H. Personalized brachytherapy dose reconstruction using deep learning. *Comput Biol Med*. 2021; 136: 104755.

133. Zaidi H, Xu XG. Computational anthropomorphic models of the human anatomy: the path to realistic Monte Carlo modeling in radiological sciences. *Annu Rev Biomed Eng*. 2007; 9: 471-500.

134. Bolch WE, Bouchet LG, Robertson JS, Wessels BW, Siegel JA, Howell RW, et al. MIRD pamphlet No. 17: the dosimetry of nonuniform activity distributions—radionuclide S values at the voxel level. medical internal radiation dose committee. *J Nucl Med*. 1999; 40: 11-36.

135. Johnson PB, Whalen SR, Wayson M, Juneja B, Lee C, Bolch WE. Hybrid patient-dependent phantoms covering statistical distributions of body morphometry in the U.S. adult and pediatric population. *Proc IEEE*. 2009; 97: 2060-75.

136. Akhavanallaf A, Xie T, Zaidi H. Development of a library of adult computational phantoms based on anthropometric indexes. *IEEE Trans Radiat Plasma Med Sci*. 2019; 3: 65-75.

137. Na YH, Zhang B, Zhang J, Caracappa PF, Xu XG. Deformable adult human phantoms for radiation protection dosimetry: anthropometric data representing size distributions of adult worker populations and software algorithms. *Phys Med Biol*. 2010; 55: 3789-811.

138. Xie T, Akhavanallaf A, Zaidi H. Construction of patient-specific computational models for organ dose estimation in radiological imaging. *Med Phys*. 2019; 46: 2403-11.

139. Berger MJ. Distribution of absorbed dose around point sources of electrons and beta particles in water and other media. *J Nucl Med*. 1971; Suppl 5: 23.

140. Kolbert KS, Sgouros G, Scott AM, Bronstein JE, Malane RA, Zhang J, et al. Implementation and evaluation of patient-specific three-dimensional internal dosimetry. *J Nucl Med*. 1997; 38: 301-8.

141. Giap HB, Macey DJ, Bayouth JE, Boyer AL. Validation of a dose-point kernel convolution technique for internal dosimetry. *Phys Med Biol*. 1995; 40: 365-81.

142. Zaidi H. Relevance of accurate monte carlo modeling in nuclear medical imaging. *Med Phys*. 1999; 26: 574-608.

143. Gardin I, Bouchet LG, Assie K, Caron J, Lisbona A, Ferrer L, et al. Voxeldoes: a computer program for 3-D dose calculation in therapeutic nuclear medicine. *Cancer Biother Radiopharm*. 2003; 18: 109-15.

144. Papadimitroulas P, Loudos G, Nikiforidis GC, Kagadis GC. A dose point kernel database using GATE monte carlo simulation toolkit for nuclear medicine applications: comparison with other monte carlo codes. *Med Phys*. 2012; 39: 5238-47.

145. Besemer AE, Yang YM, Grudzinski JJ, Hall LT, Bednarz BP. Development and validation of RAPID: a patient-specific monte carlo three-dimensional internal dosimetry platform. *Cancer Biother Radiopharm*. 2018; 33: 155-65.

146. Ljungberg M, Sjogren Gleisner K. 3-D image-based dosimetry in radionuclide therapy. *IEEE Trans Radiat Plasma Med Sci*. 2018; 2: 527-40.

147. Mok GSP, Dewaraja YK. Recent advances in voxel-based targeted radionuclide therapy dosimetry. *Quant Imaging Med Surg*. 2021; 11: 483-9.

148. Dieudonne A, Hobbs RF, Lebtahi R, Maurel F, Baechler S, Wahl RL, et al. Study of the impact of tissue density heterogeneities on 3-dimensional abdominal dosimetry: comparison between dose kernel convolution and direct monte carlo methods. *J Nucl Med*. 2013; 54: 236-43.

149. Loudos G, Tsougos I, Boukis S, Karakatsanis N, Georgoulias P, Theodorou K, et al. A radionuclide dosimetry toolkit based on material-specific monte carlo dose kernels. *Nucl Med Commun*. 2009; 30: 504-12.

150. Khazaei Moghadam M, Kamali Asl A, Geramifar P, Zaidi H. Evaluating the application of tissue-specific dose kernels instead of water dose kernels in internal dosimetry: a monte carlo study. *Cancer Biother Radiopharm*. 2016; 31: 367-79.

151. Lee MS, Kim JH, Paeng JC, Kang KW, Jeong JM, Lee DS, et al. Whole-body voxel-based personalized dosimetry: the multiple voxel s-value approach for heterogeneous media with nonuniform activity distributions. *J Nucl Med*. 2018; 59: 1133-9.

152. Arabi H, Zaidi H. Deep learning-guided estimation of attenuation correction factors from time-of-flight PET emission data. *Med Image Anal*. 2020; 64: 101718.

153. Xiang H, Lim H, Fessler JA, Dewaraja YK. A deep neural network for fast and accurate scatter estimation in quantitative SPECT/CT under challenging scatter conditions. *Eur J Nucl Med Mol Imaging*. 2020; 47: 2956-67.

154. Dong X, Lei Y, Wang T, Higgins K, Liu T, Curran WJ, et al. Deep learning-based attenuation correction in the absence of structural information for whole-body positron emission tomography imaging. *Phys Med Biol*. 2020; 65: 055011.

155. Mostafapour S, Gholamiankhah F, Dadgar H, Arabi H, Zaidi H. Feasibility of deep learning-guided attenuation and scatter correction of whole-body 68Ga-PSMA PET studies in the image domain. *Clin Nucl Med*. 2021; 46: 609-15.

156. Arabi H, Zaidi H. MRI-guided attenuation correction in torso PET/MRI: assessment of segmentation-, atlas-, and deep learning-based approaches in the presence of outliers. *Magn Reson Med*. 2022; 87: 686-701.

157. Arabi H, Zaidi H. Truncation compensation and metallic dental implant artefact reduction in PET/MRI attenuation correction using deep learning-based object completion. *Phys Med Biol*. 2020; 65: 195002.

158. Sanaat A, Arabi H, Mainta I, Garibotti V, Zaidi H. Projection space implementation of deep learning-guided low-dose brain PET imaging improves performance over implementation in image space. *J Nucl Med*. 2020; 61: 1388-96.

159. Zaharchuk G. Next generation research applications for hybrid PET/MR and PET/CT imaging using deep learning. *Eur J Nucl Med Mol Imaging*. 2019; 46: 2700-7.

160. Aghakhan Olia N, Kamali-Asl A, Hariri Tabrizi S, Geramifar P, Sheikzadeh P, Farzanefar S, et al. Deep learning-based denoising of low-dose SPECT myocardial perfusion images: quantitative assessment and clinical performance. *Eur J Nucl Med Mol Imaging*. 2022; 49: 1508-22.

161. Sanaei B, Faghihi R, Arabi H. Employing multiple low-dose PET images (at different dose levels) as prior knowledge to predict standard-dose PET images. *J Digit Imaging*. 2023; 36: 1588-96.

162. Azimi MS, Kamali-Asl A, Ay MR, Zeraatkar N, Hosseini MS, Sanaat A, et al. Deep learning-based partial volume correction in standard and low-dose positron emission tomography-computed tomography imaging. *Quant Imaging Med Surg*. 2024; 14: 2146-64.

163. Seo H, Badiee Khuzani M, Vasudevan V, Huang C, Ren H, Xiao R, et al. Machine learning techniques for biomedical image segmentation: An overview of technical aspects and introduction to state-of-art applications. *Med Phys*. 2020; 47: 148-67.

164. Boehringer AS, Sanaat A, Arabi H, Zaidi H. An active learning approach to train a deep learning algorithm for tumor segmentation from brain MR images. *Insights Imaging*. 2023; 14: 141.

165. Safarian A, Mirshahvalad SA, Nasrollahi H, Jung T, Pirich C, Arabi H, et al. Impact of [(18)F]FDG PET/CT radiomics and artificial intelligence in clinical decision making in lung cancer: its current role. *Semin Nucl Med*. 2025; March 5.

166. Mardani M, Dong P, Xing L. Deep-learning based prediction of achievable dose for personalizing inverse treatment planning. *Int J Radiat Oncol Biol Phys*. 2016; 96: E419-E20.

167. Nguyen D, Long T, Jia X, Lu W, Gu X, Iqbal Z, et al. A feasibility study for predicting optimal radiation therapy dose distributions of prostate cancer patients from patient anatomy using deep learning. *Sci Rep*. 2019; 9: 1076.

168. Ma M, M KB, Vasudevan V, Xing L, Yang Y. Dose distribution prediction in isodose feature-preserving voxelization domain using deep convolutional neural network. *Med Phys*. 2019; 46: 2978-87.

169. Kearney V, Chan JW, Haaf S, Descovich M, Solberg TD. DoseNet: a volumetric dose prediction algorithm using 3D fully-convolutional neural networks. *Phys Med Biol*. 2018; 63: 235022.

170. Jarrett D, Stride E, Vallis K, Gooding MJ. Applications and limitations of machine learning in radiation oncology. *Br J Radiol*. 2019; 92: 20190001.

171. Andreo P. Monte Carlo simulations in radiotherapy dosimetry. *Radiat Oncol*. 2018; 13: 121.

172. Lee MS, Hwang D, Kim JH, Lee JS. Deep-dose: a voxel dose estimation method using deep convolutional neural network for personalized internal dosimetry. *Sci Rep*. 2019; 9: 10308.

173. Gotz TI, Schmidkonz C, Chen S, Al-Baddai S, Kuwert T, Lang EW. A deep learning approach to radiation dose estimation. *Phys Med Biol*. 2020; 65: 035007.

174. Eckerman K, Endo A. ICRP Publication 107. Nuclear decay data for dosimetric calculations. *Ann ICRP*. 2008; 38: 7-96.

175. Chhabra A, Thakur ML. Theragnostic radionuclide pairs for prostate cancer management: (64)Cu/(67)Cu, can be a budding hot duo. *Biomedicines*. 2022; 10: 2787.

176. Vaidyanathan G, Zalutsky MR. Targeted therapy using alpha emitters. *Phys Med Biol*. 1996; 41: 1915-31.

177. Deshayes E, Roumiguié M, Thibault C, Beuzeboc P, Cachin F, Hennequin C, et al. Radium 223 dichloride for prostate cancer treatment. *Drug Des Devel Ther*. 2017; 11: 2643-51.

178. Du Y, Carrio I, De Vincentis G, Fanti S, Ilhan H, Mommsen C, et al. Practical recommendations for radium-223 treatment of metastatic castration-resistant prostate cancer. *Eur J Nucl Med Mol Imaging*. 2017; 44: 1671-8.

179. Abbott A, Sakellis CG, Andersen E, Kuzuhara Y, Gilbert L, Boyle K, et al. Guidance on (177)Lu-DOTATATE peptide receptor radionuclide therapy from the experience of a single nuclear medicine division. *J Nucl Med Technol*. 2018; 46: 237-44.

180. Kratochwil C, Haberkorn U, Giesel FL. Radionuclide therapy of metastatic prostate cancer. *Semin Nucl Med*. 2019; 49: 313-25.

181. Kean SJ. Lutetium Lu 177 pipevacetide tetraxetan: first approval. *Mol Diagn Ther*. 2022; 26: 467-75.

182. Sanli Y, Simsek DH, Sanli O, Subramaniam RM, Kendi AT. (177)Lu-PSMA therapy in metastatic castration-resistant prostate cancer. *Biomedicines*. 2021; 385(12):1091-1103.

183. Banerjee S, Pillai MR, Knapp FF. Lutetium-177 therapeutic radiopharmaceuticals: linking chemistry, radiochemistry, and practical applications. *Chem Rev*. 2015; 115: 2934-74.

184. Groener D, Nguyen CT, Baumgarten J, Bockisch B, Davis K, Happel C, et al. Hematologic safety of (177)Lu-PSMA-617 radioligand therapy in patients with metastatic castration-resistant prostate cancer. *EJNMMI Res*. 2021; 11: 61.

185. Kind F, Michalski K, Yousefzadeh-Nowshahr E, Meyer PT, Mix M, Ruf J. Bone marrow impairment during early [(177)Lu]PSMA-617 radioligand therapy: haematotoxicity or tumour progression? *EJNMMI Res.* 2022; 12: 20.

186. Gudenkauf LM, Chavez MN, Maconi ML, Geiss C, Seyedroudbari A, Thin P, et al. Developing a patient-reported outcome measure for radionuclide therapy for prostate cancer. *J Nucl Med.* 2023; 64: 869-72.

187. Demir M, Abuqbeitah M, Uslu-Besli L, Yildirim O, Yeyin N, Cavdar I, et al. Evaluation of radiation safety in (177)Lu-PSMA therapy and development of outpatient treatment protocol. *J Radiol Prot.* 2016; 36: 269-78.

188. Mittra ES. Neuroendocrine tumor therapy: (177)Lu-DOTATATE. *AJR Am J Roentgenol.* 2018; 211: 278-85.

189. Kurth J, Krause BJ, Schwarzenbock SM, Stegger L, Schafers M, Rahbar K. External radiation exposure, excretion, and effective half-life in (177)Lu-PSMA-targeted therapies. *EJNMMI Res.* 2018; 8: 32.

190. Sanli Y, Simsek DH, Sanli Ö, Subramaniam RM, Kendi AT. 177Lu-PSMA therapy in metastatic castration-resistant prostate cancer. *Biomedicines.* 2021; 9: 430.

191. Mikhail-Lette M, Cordero L, Lievens Y, Al-Ibraheem A, Urbain JL, Chera B, et al. Six country vignettes: strengthening radiotherapy and theranostics. *J Cancer Policy.* 2024; 40: 100471.

192. Al-Ibraheem A. Theranostics in developing countries: addressing challenges and potentials from training to practice. *World J Nucl Med.* 2023; 22: 171-3.

193. Al-Ibraheem A, Zimmermann R, Abdulkadir AS, Herrmann K. Radiotheranostics global market and future developments. *Semin Nucl Med.* 2024; 54:622-633.